

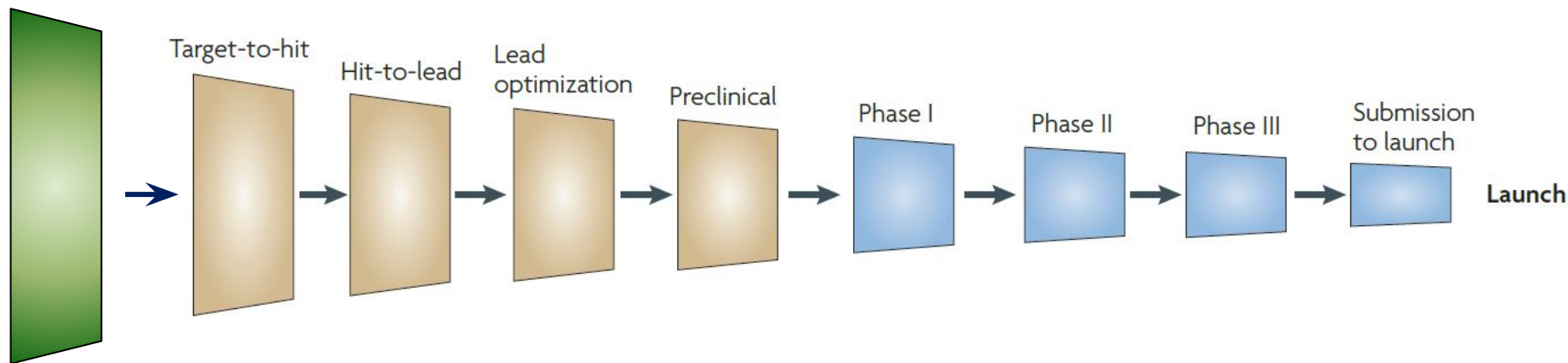
What can we do if there are no good targets

*Mathematical and Computational Biology in Drug Discovery
(MCBDD) Module II*

*Dr. Jitao David Zhang
March-April 2021*

The linear view of drug discovery builds on target-based approaches

Target identification & assessment

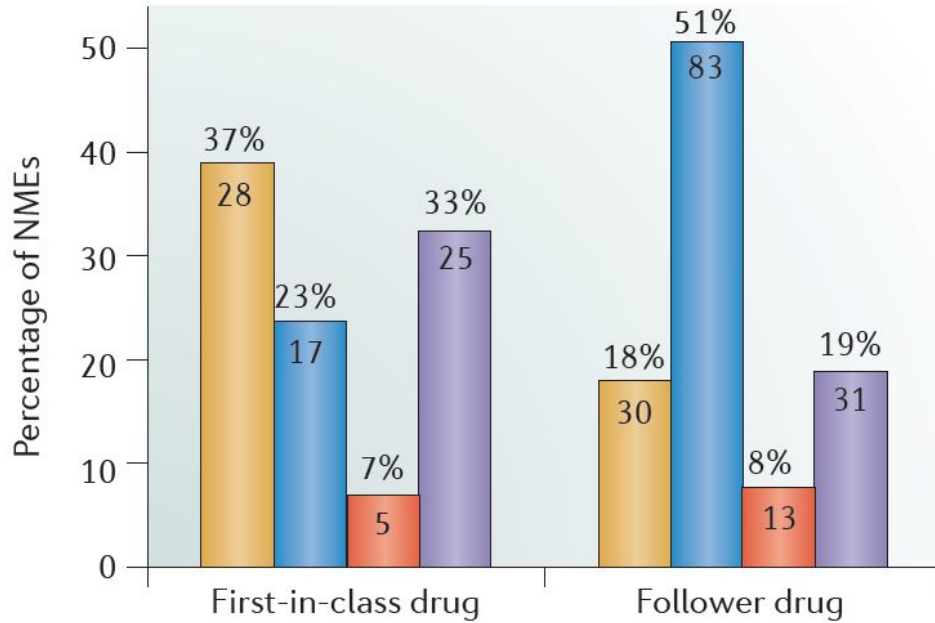
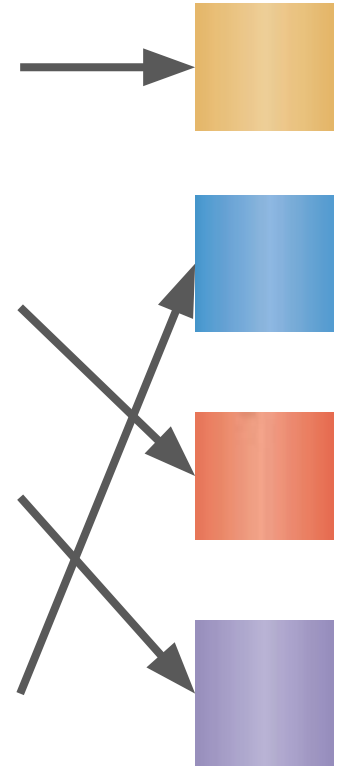


Five strategies when no good target is found

1. Phenotypic drug discovery
2. Natural products
3. Biologics
4. Interaction-based (multispecific) drug discovery
5. Drug repurposing or combination studies

Connect the lines!

- Phenotypic screening
- Modified natural products
- Biologics
- Target-based screening



Phenotypic screenings by agent and readout

Agent

High-throughput screening
libraries ($\geq 10^6$ molecules)

Genetic libraries ($\sim 10^4$)

Natural products and chemo-
genomic libraries ($\sim 10^3$)

Custom libraries ($\sim 10^0 - 10^2$)

Feasibility

Reporters

Gene
expression

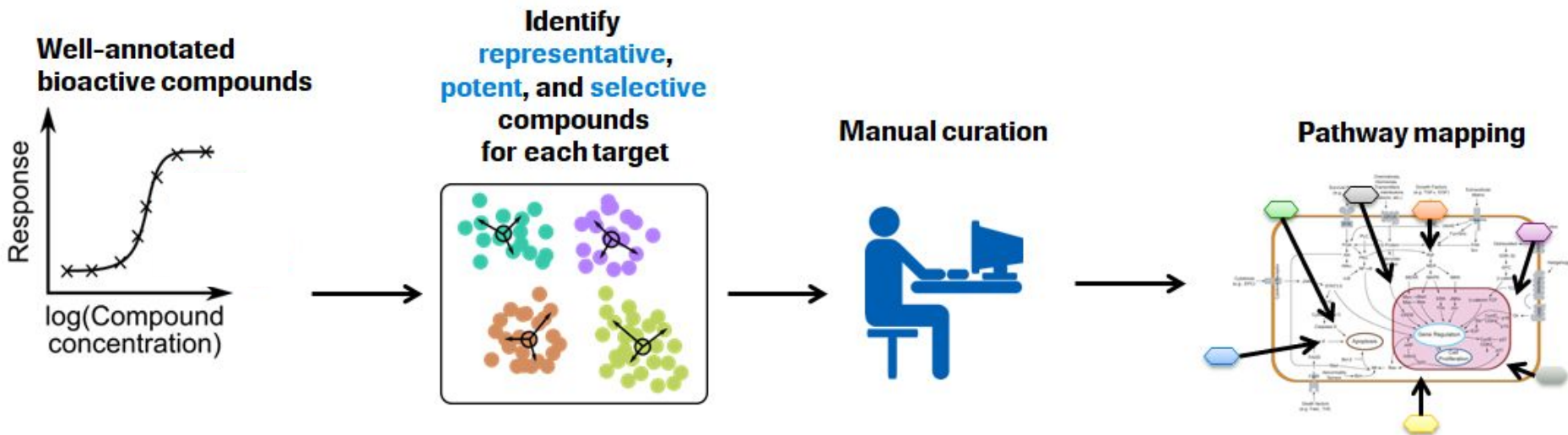
Cellular
morphology

Organ/tissue
phenotype

Organism
phenotype

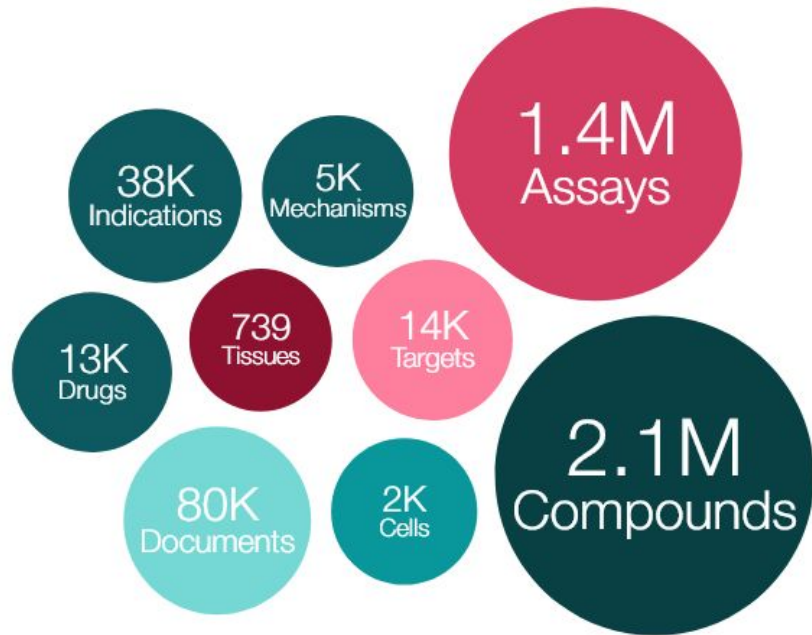
Readout

The Small-molecule Pathway Research Kit (SPARK)



The ChEMBL database

- An example of query: [aspirin](#).
- Systematic and programmatic accession via [ChEMBL API](#) ([source code](#)).
- We can use **dose-response data** to annotate the *triplets* of compound, assay activity, and targets.

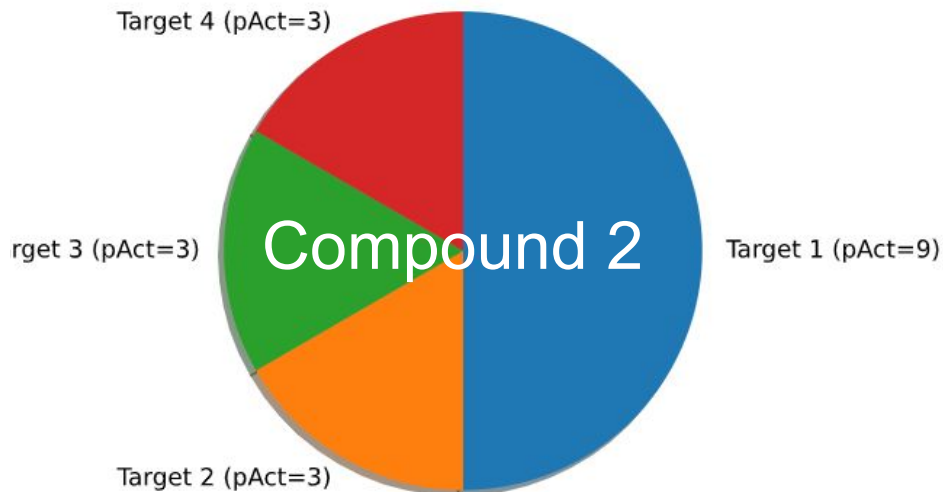
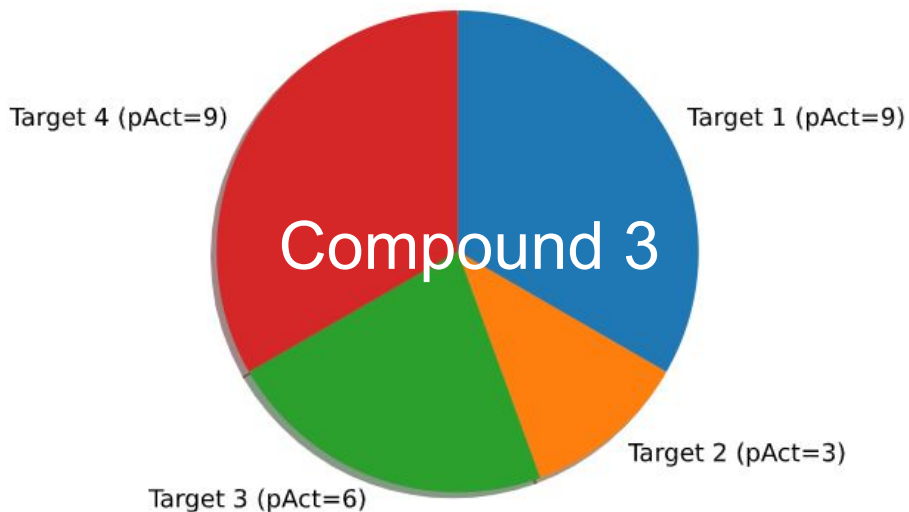
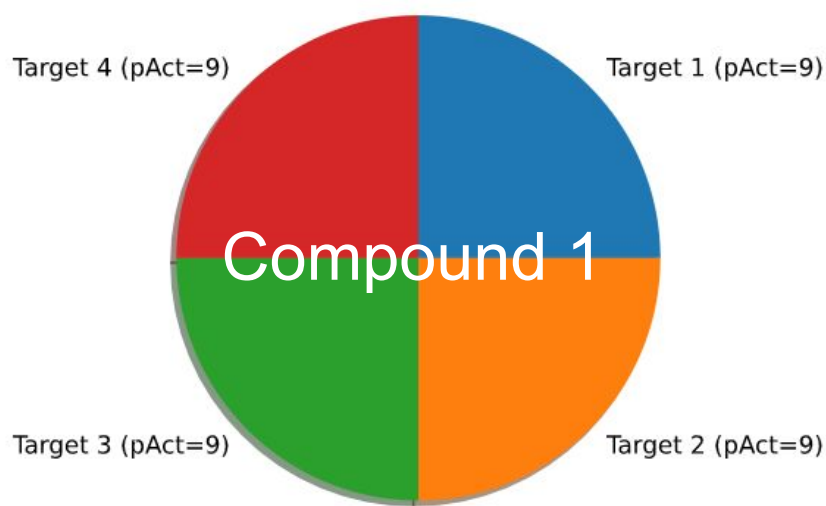


March 2021

Discussion

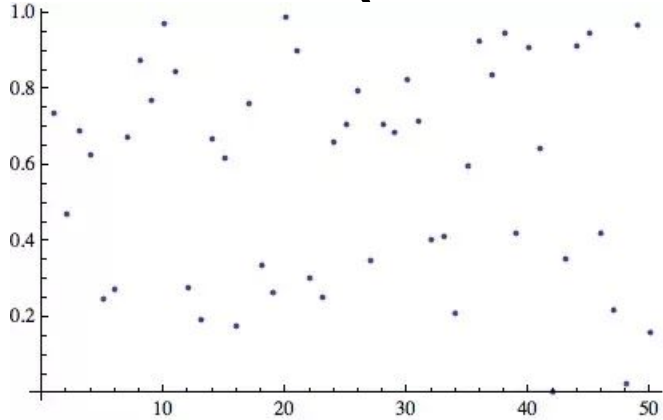
1. Why do we care selecting *representative, potent, and selective* compounds for each target?
2. How to define following terms mathematically ...
 - a. Representativity?
 - b. Potency?
 - c. Selectivity?

A toy example about how to quantify a compound's potency and selectivity

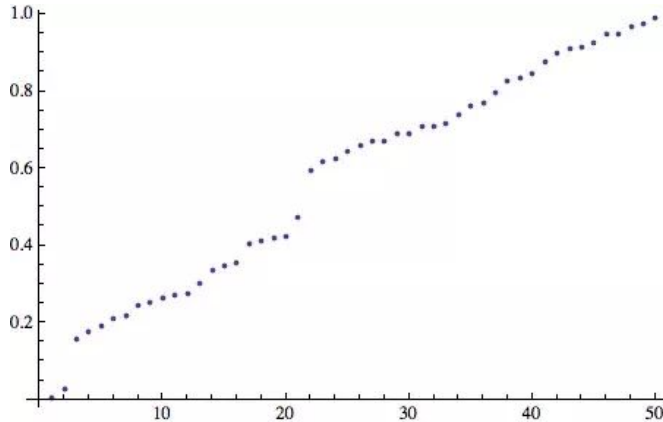


The Gini Index (a.k.a. Gini Coefficient)

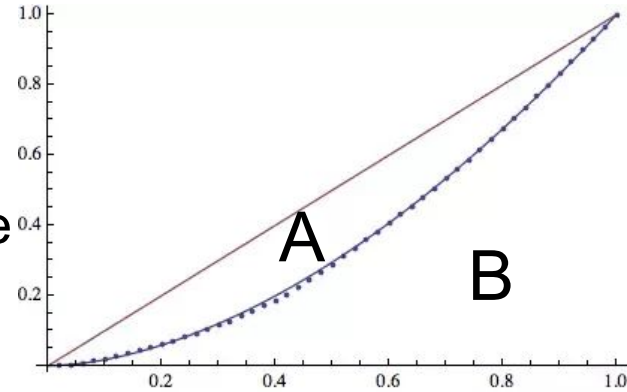
A random
vector of
50 values



Sorted
from low
to high



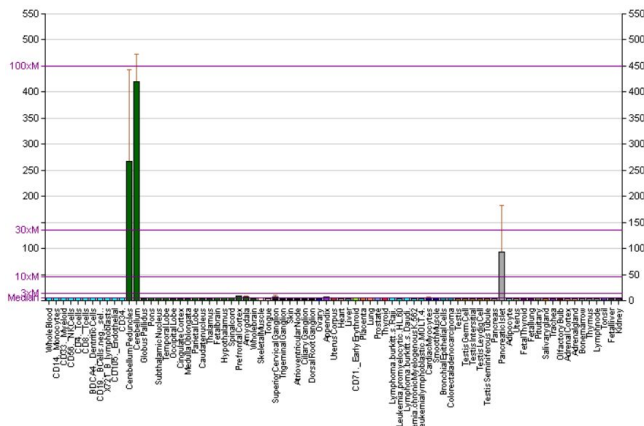
The Gini
Index is
calculated
based on the
cumulative
distribution



$$G=A/(A+B)$$

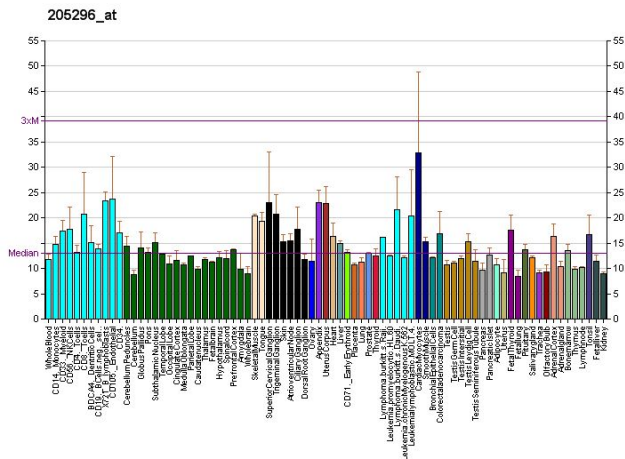
The Gini Index quantifies inequality/ selectivity

NEUROD1

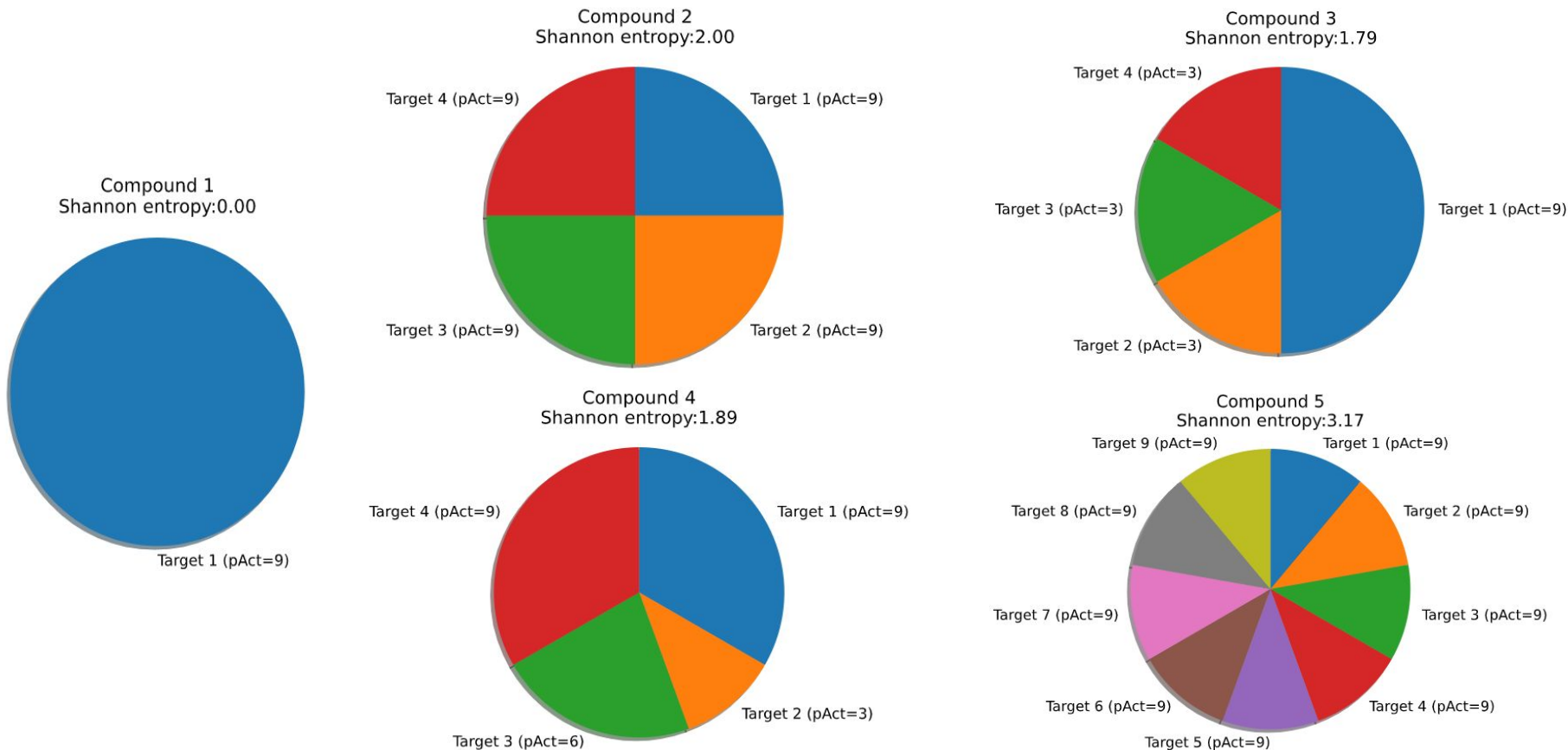


The Gini Index of expression of *NEUROD1* across tissues is near 1, whereas that of *RBL1* is near 0.

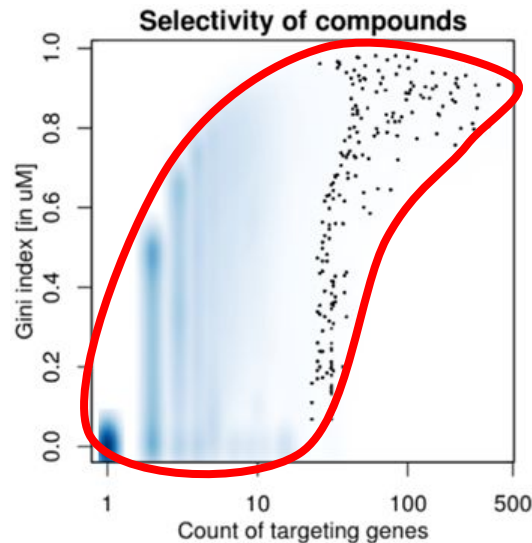
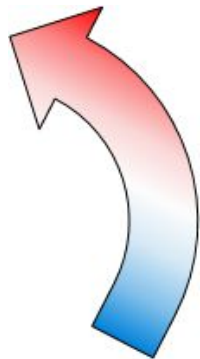
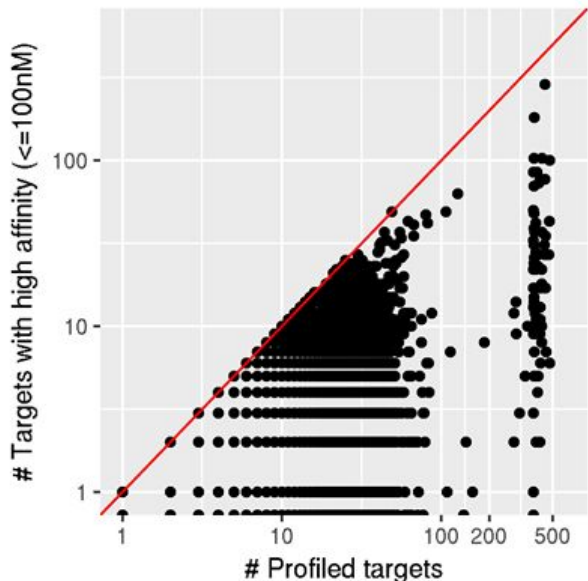
RBL1



An alternative metric: Shannon's Entropy



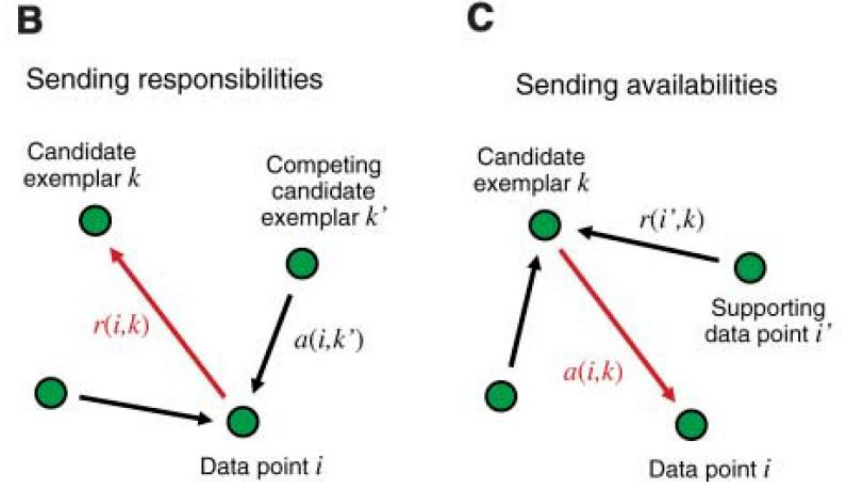
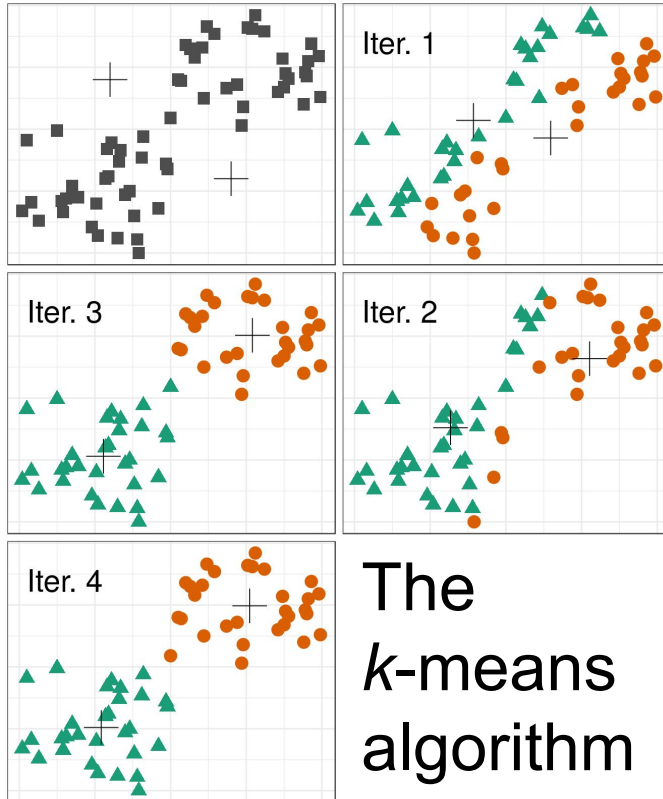
Count of targets and selectivity of ChEMBL molecules



With some exceptions, most compounds are profiled against <100 targets. We distinguish between specific and pleiotropic compounds.

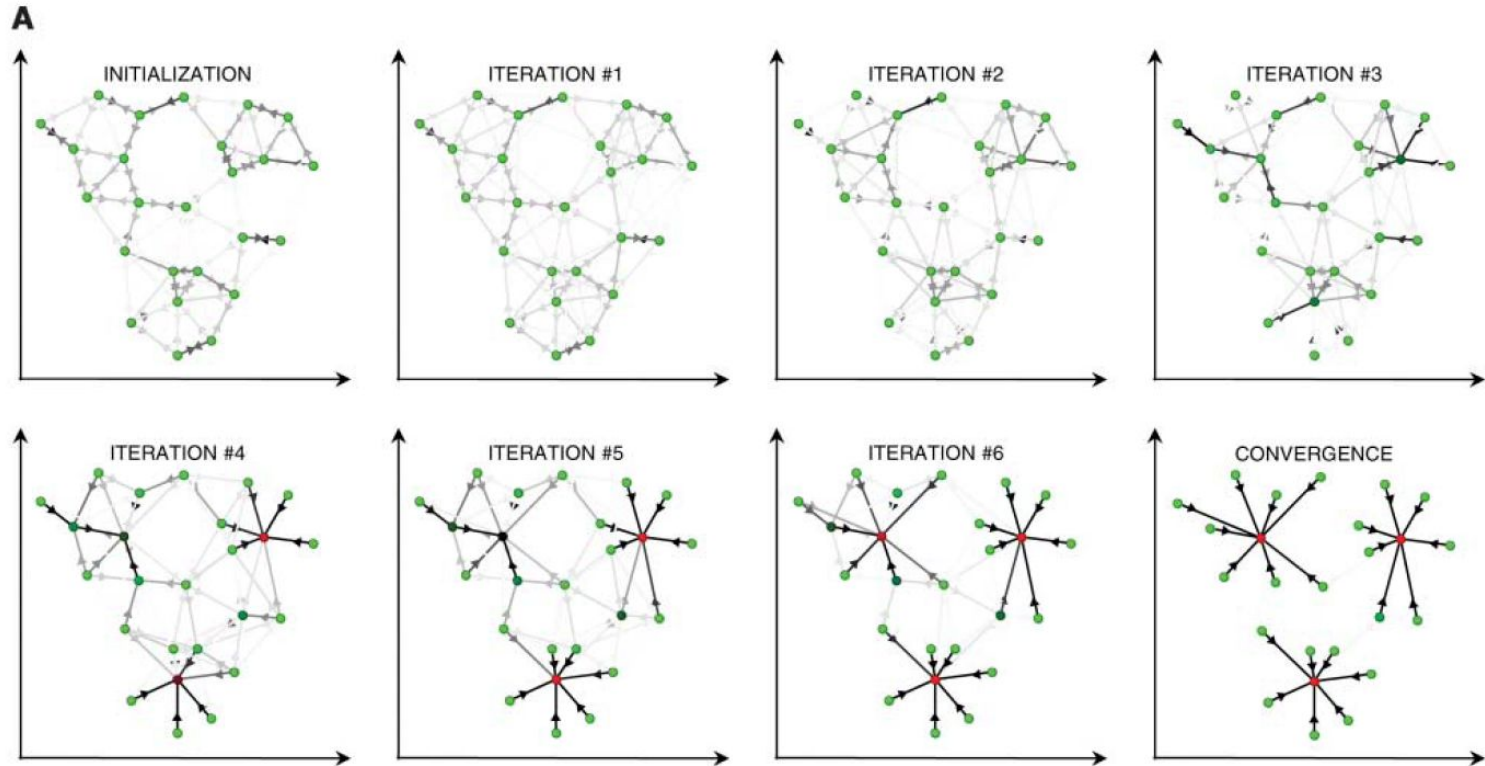
The **shark-fin shape** curve suggests that frequently profiled compounds tend to be more selective (and *vice versa*).

Unsupervised clustering



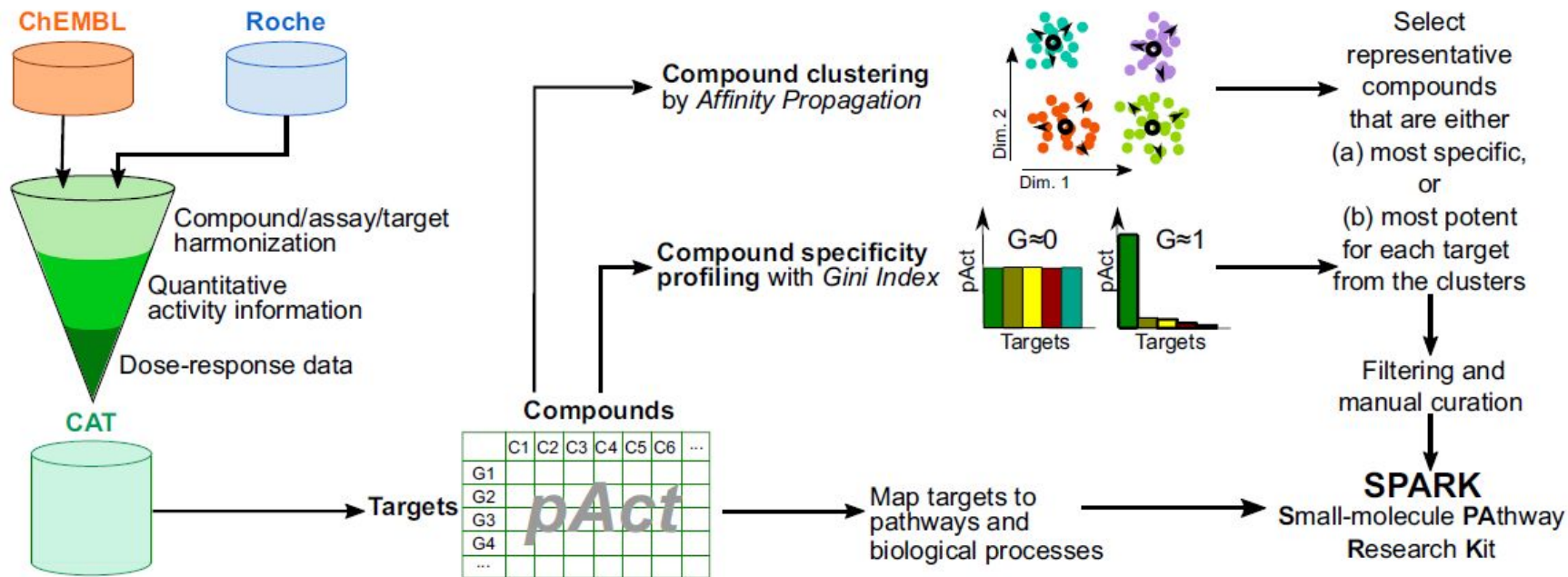
Affinity Propagation updates **responsibilities** and **availabilities** iteratively

Affinity Propagation in action



[A movie of iterations](#)

Construction of SPARK in detail



Harmonization

... of public and Roche internal data

Machine learning

... to select compounds

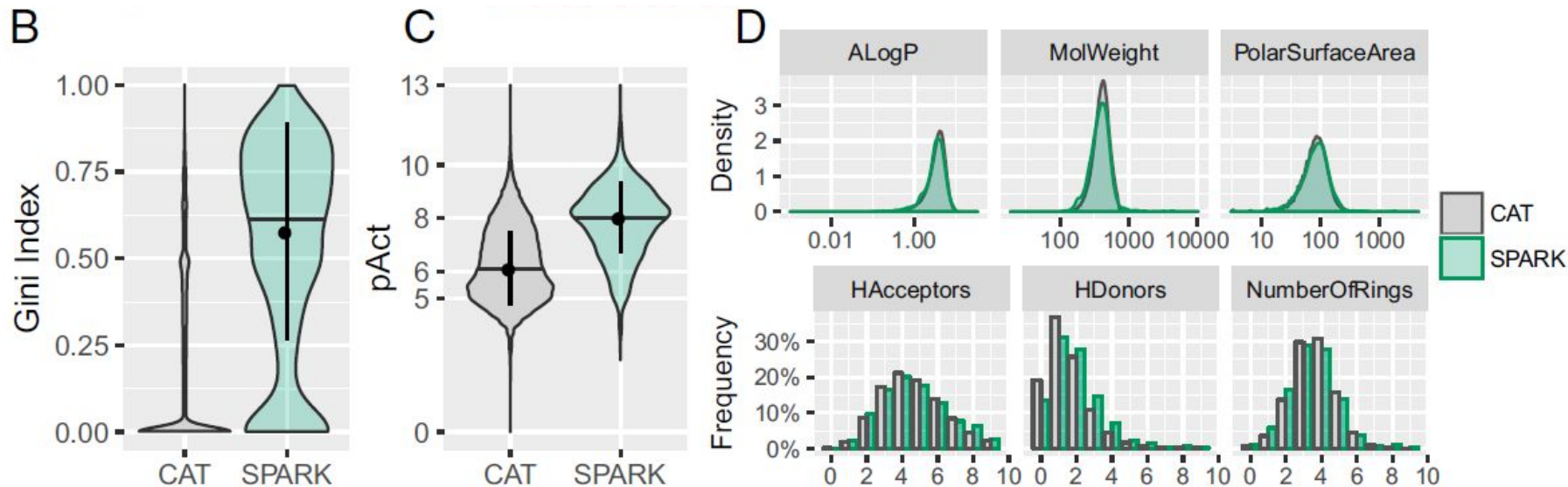
Pathways

... mapped to compounds

Curation

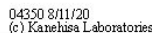
... to enrich quality compounds

SPARK covers the chemical space evenly with representative, potent, and specific compounds



Roudnicky *et al.*, PNAS, 2020,
<https://www.pnas.org/content/early/2020/08/04/1911532117>

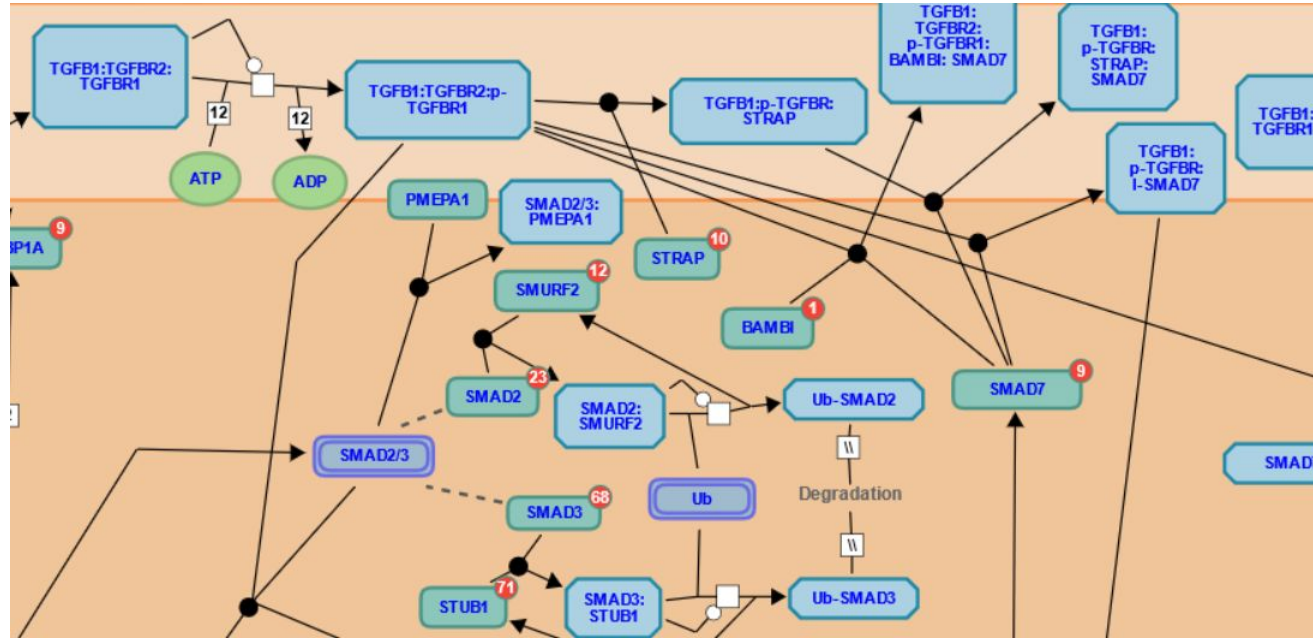
A RESTful API is available for academic use, with clients in Python and R.



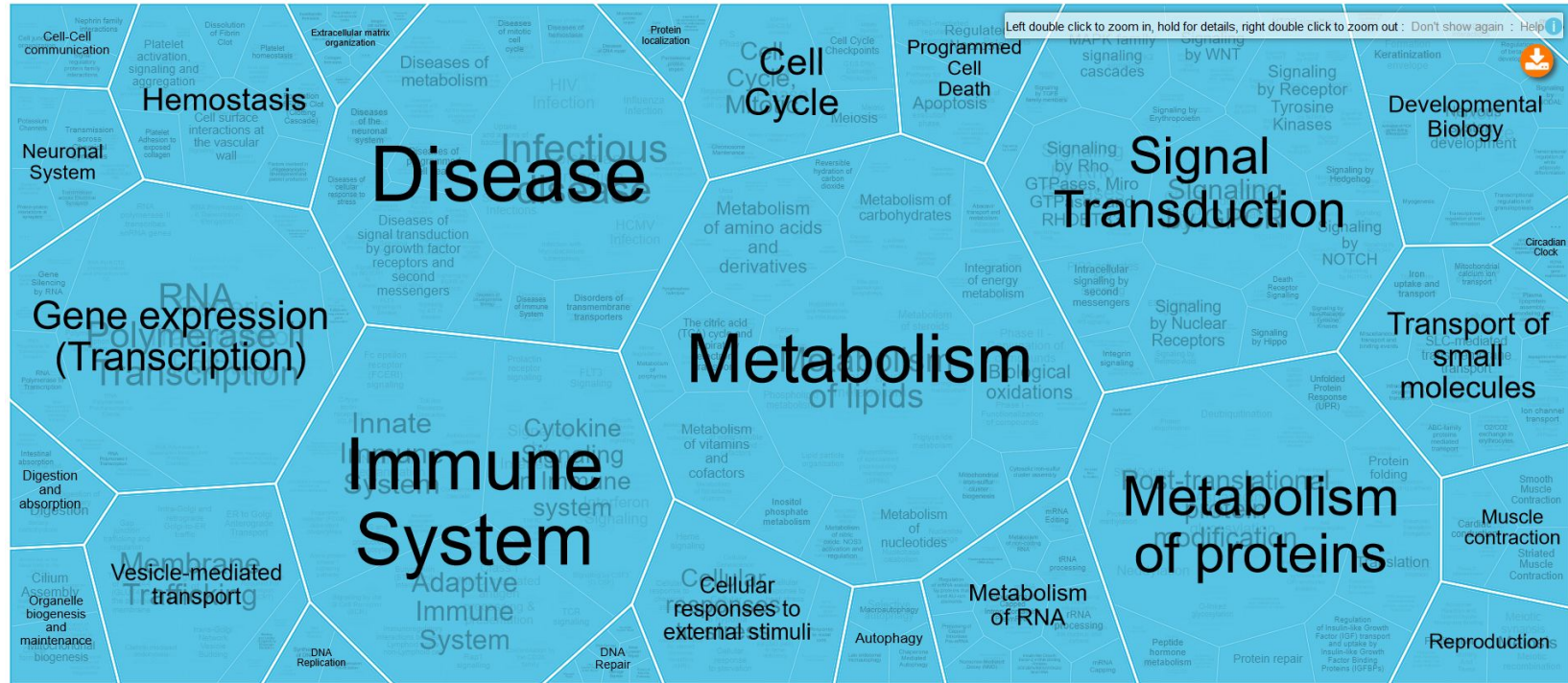
Mapping genes to biological pathways

Option 2: Reactome pathways, with the example of the TGF- β signaling pathway.

Developer's Zone
provides API and
graph database
interfaces.



Overview of pathways captured by Reactome



The Voronoi (Reacfoam) view of all pathways in Reactome

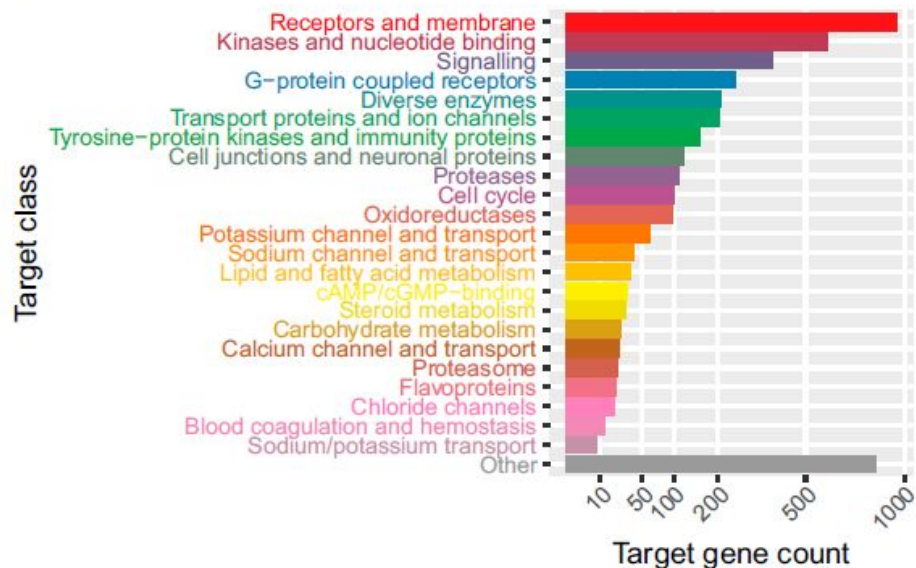
- [illegible]

- activation of protein kinase activity Source: BHF-UCL
- aging Source: Ensembl
- animal organ regeneration Source: Ensembl
- apoptotic process Source: UniProtKB
- atrioventricular valve morphogenesis Source: BHF-UCL
- blood vessel development Source: BHF-UCL
- brain development Source: BHF-UCL

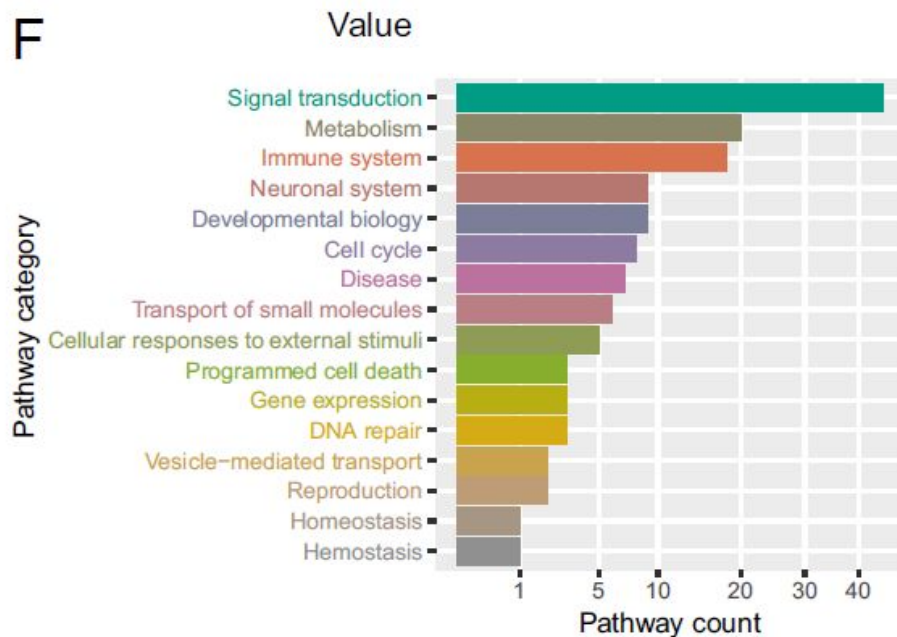
Ligand	ATP-binding, Magnesium, Manganese, Metal-binding, Nucleotide-binding
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SPARK covers the target space evenly with representative, potent, and specific compounds

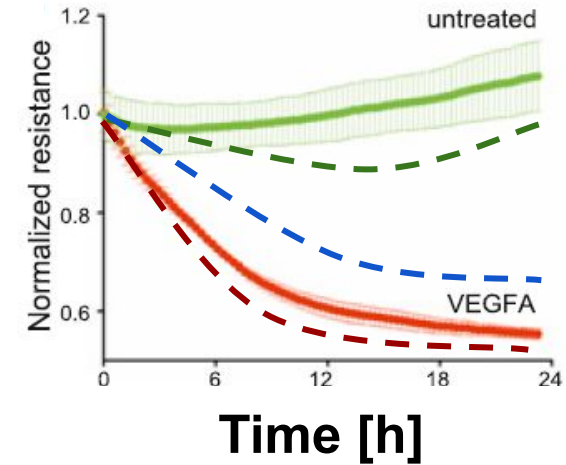
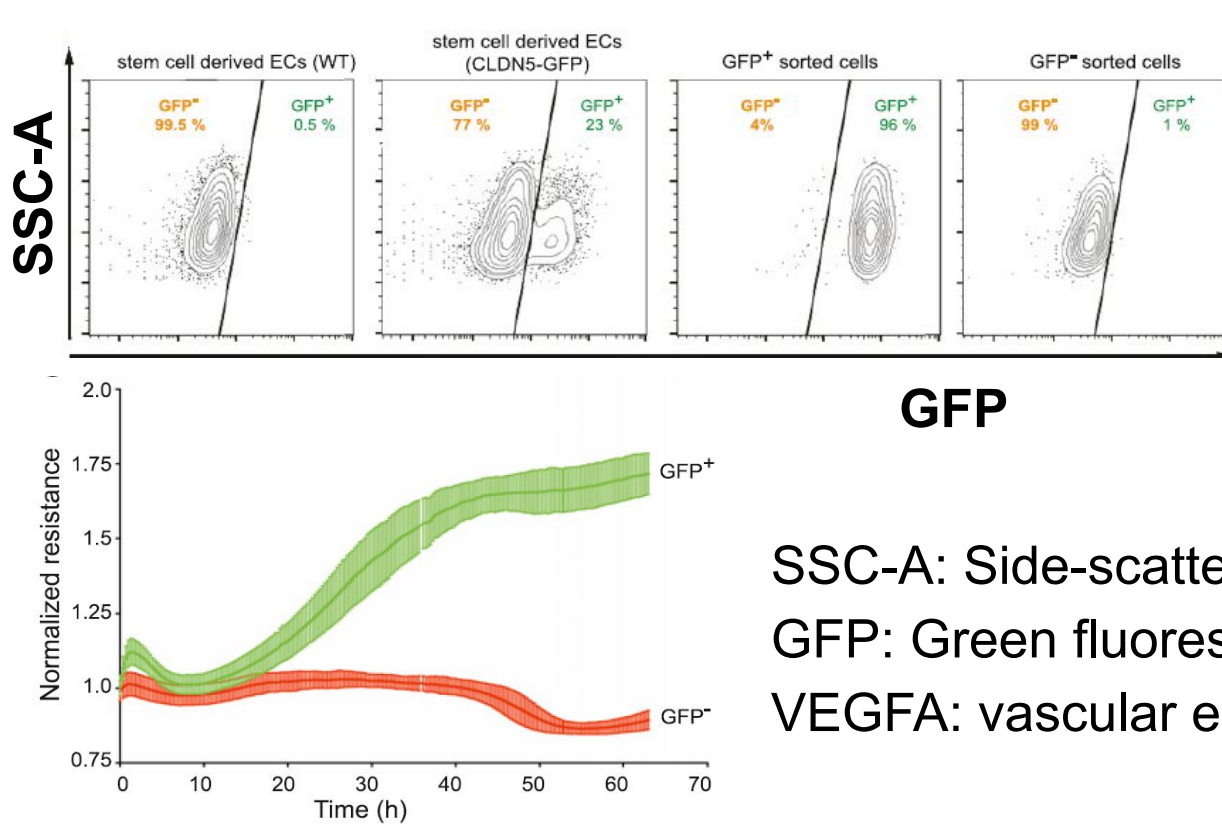
E



F

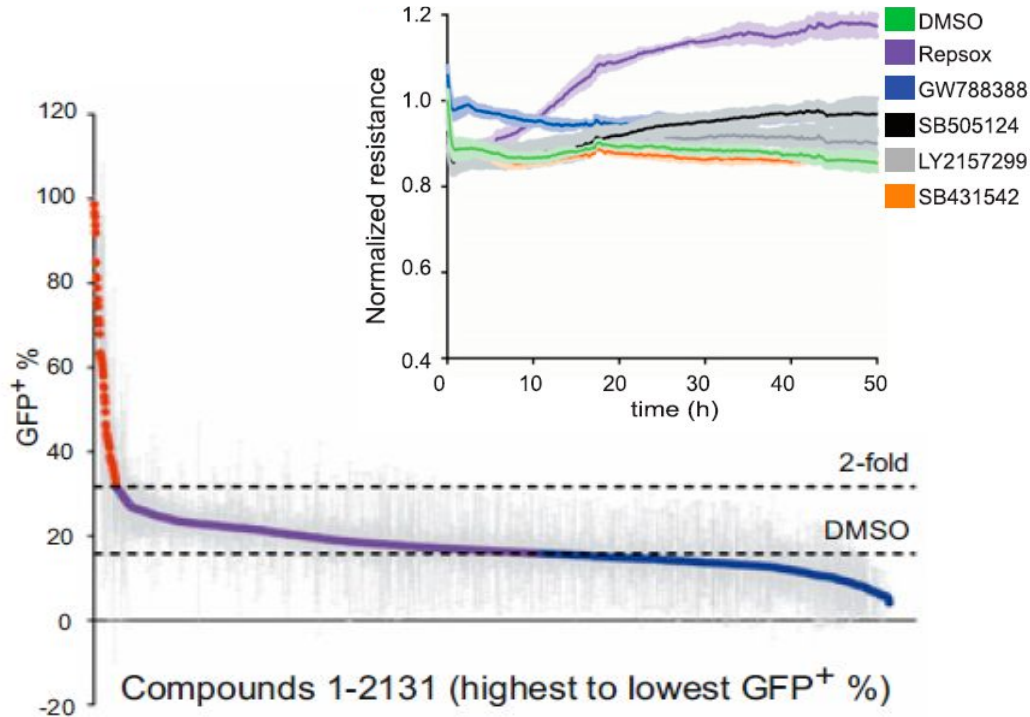


Screening with stem-cell-derived endothelial cells with a reporter added by genome editing



SSC-A: Side-scatter area of flow cytometry;
GFP: Green fluorescent protein;
VEGFA: vascular endothelial growth factor A

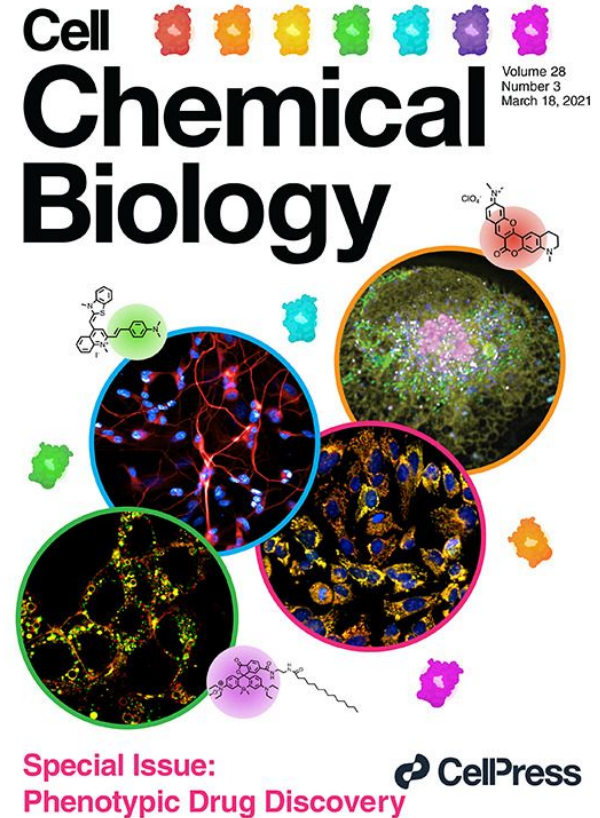
Compounds targeting the TGF- β pathway such as RepSox modulates endothelial cells



Further *in vitro* and *in vivo* experiments establish RepSox as a tool compound modulating retinopathy.

Conclusions about chemogenomic library

- Phenotypic drug discovery can lead to first-in-class drugs with novel mechanisms;
- Unsupervised machine learning and data modelling contribute to build chemogenomic libraries;
- We can link drug candidates via targets to biological pathways and processes.

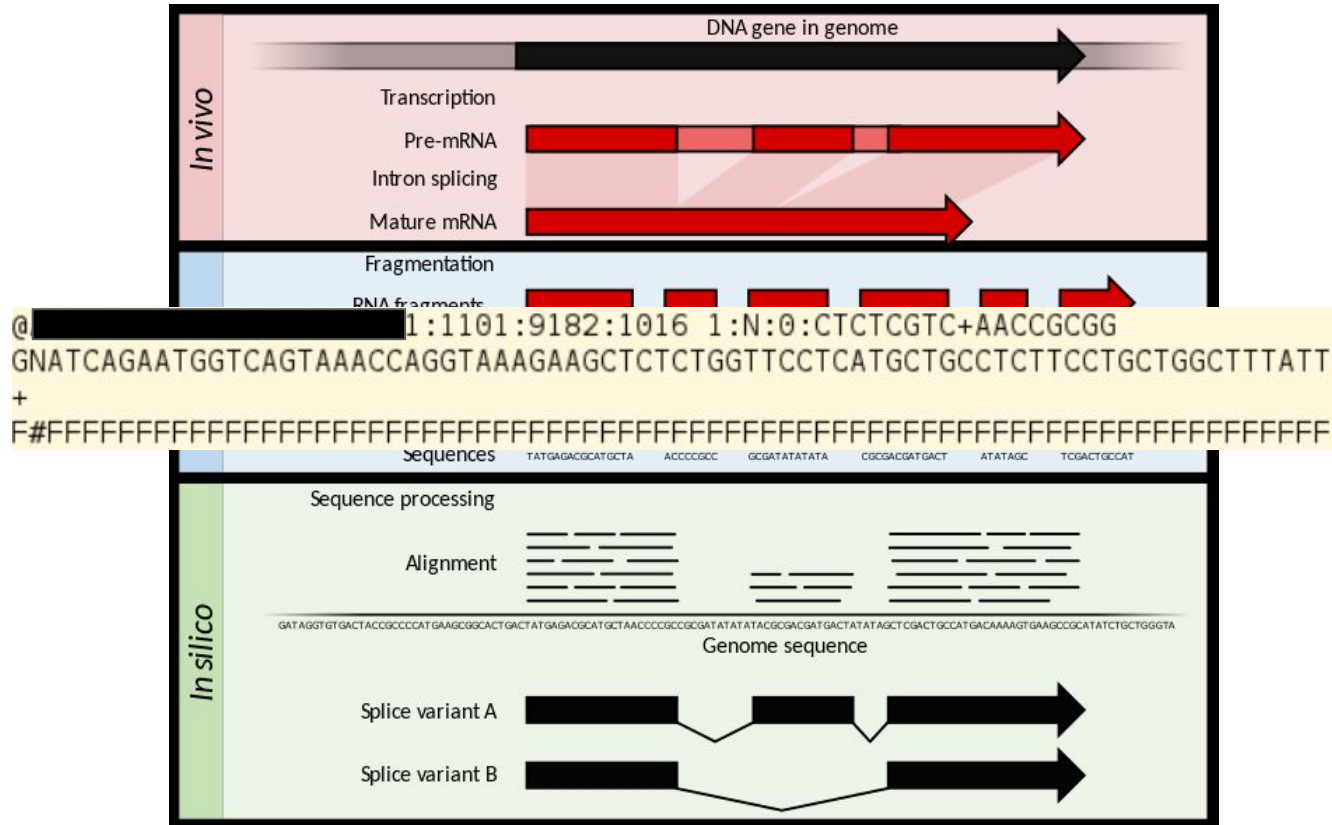


Offline activities of Module II

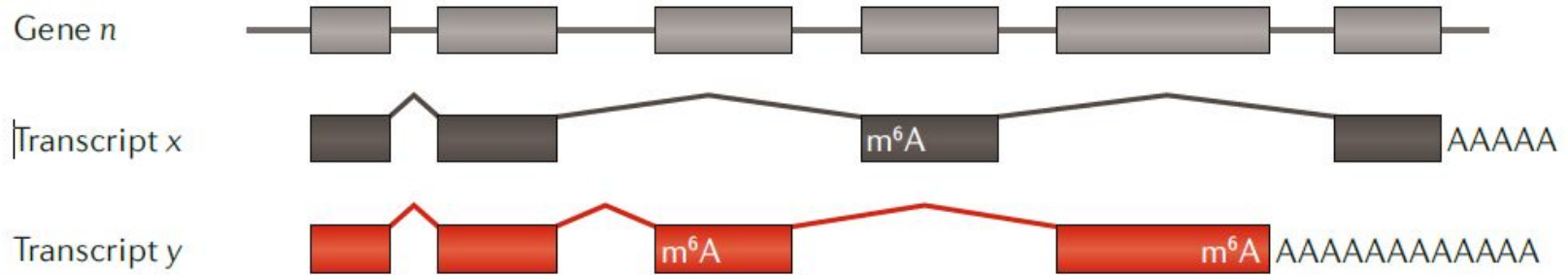
Please use your favourite programming language (shell scripts, python, R, for instance) and APIs (Application Programming Interfaces) of databases to perform following operations. Submit your code.

1. Retrieve all approved drugs from the ChEMBL database, sort them by approval year and name ([a Python example is here](#); documentations of the ChEMBL API can be found [here](#));
2. For each approved drug **since 2011** that you identified in step (1), retrieve a list of UniProt accession numbers, namely protein targets associated with the drug;
3. For each protein with a UniProt accession number that you identified in step (2), retrieve UniProt keywords associated with it. [You can use the UniProt API, documented here](#). [Python](#) and [R](#) clients are also available.

Transcriptome profiling by RNA sequencing



Transcriptome profiling by RNA sequencing



Ambiguous
to exon



Unambiguous
to exon



Ambiguous
to isoform



Unambiguous
to isoform

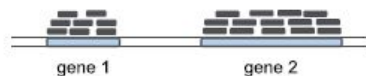


Read Mapping

Count collection

Normalization by library size

Differential Gene Expression Analysis



	sample A1	sample A2	sample B1	sample B2
gene 1	8	10	100	200
gene 2	14	15	15	40
gene 3	33	40	35	70
...
gene N	100	120	105	220

	sample A1	sample A2	sample B1	sample B2
gene 1	8	10	100	200
gene 2	14	15	115	40
gene 3	33	40	35	70
...
gene N	100	120	105	220

Tot. reads:
5 millions

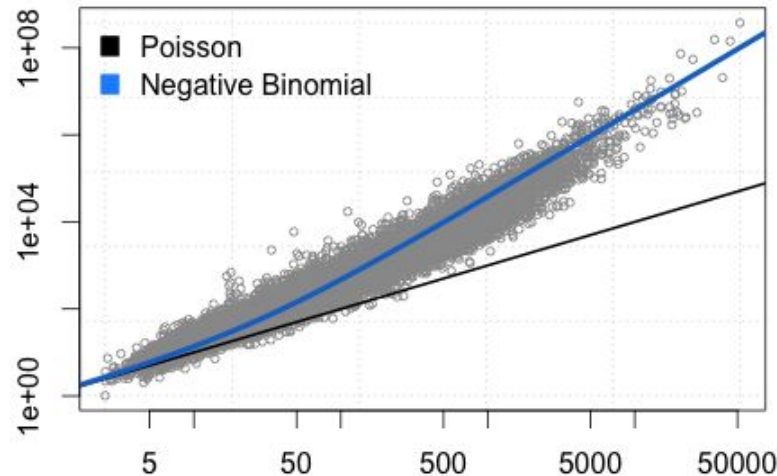
Tot. reads:
10 millions

	sample A1	sample A2	sample B1	sample B2
gene 1	0.16	0.20	2.00	2.00
gene 2	0.28	0.30	0.30	0.40
gene 3	0.66	0.80	0.70	0.70
...
gene N	2.00	2.40	2.10	2.20

Differential gene expression



Pooled gene-level variance (log10 scale)



Mean gene expression level (log10 scale)

Tools: *edgeR* and *DESeq2*

Interpret differential gene expression data with gene-set enrichment analysis

Reactome pathways

Gene Ontology

UniProt Keywords

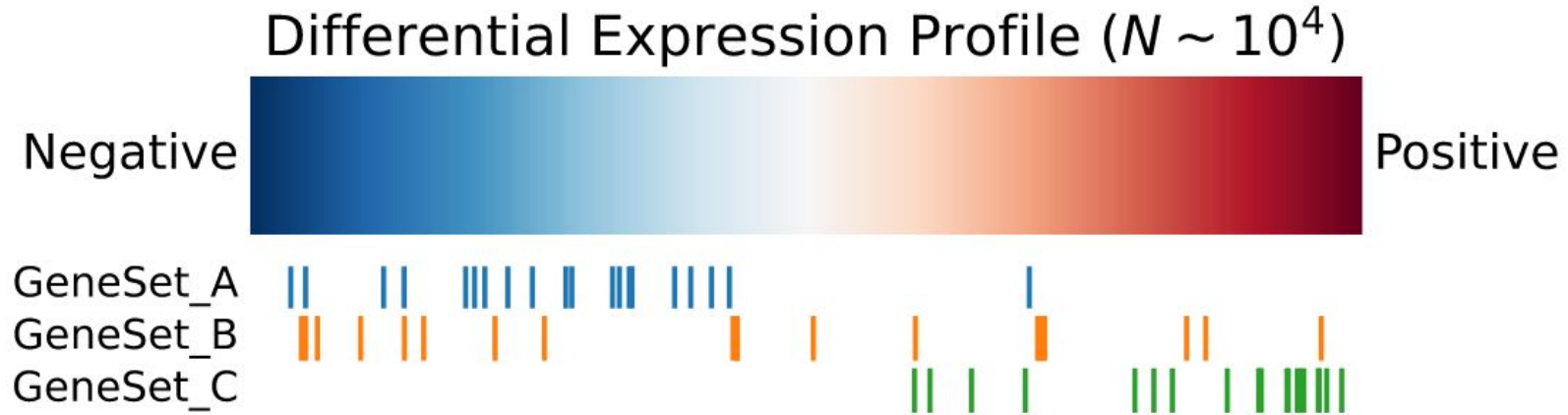
Literature

Gene (N~10 ⁴)	G ₁	G ₂	G ₃	G ₄	G ₅	...	G _{N-3}	G _{N-2}	G _{N-1}	G _N
Change (log2)	3.0	2.8	2.5	1.5	1.2	...	-0.8	-1.2	-1.5	-2.2

Differential gene expression results

Gene-set Enrichment Analysis Methods

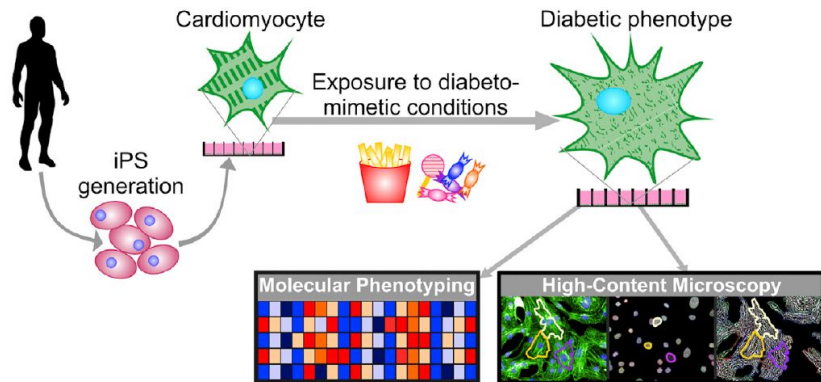
Gene-set enrichment analysis



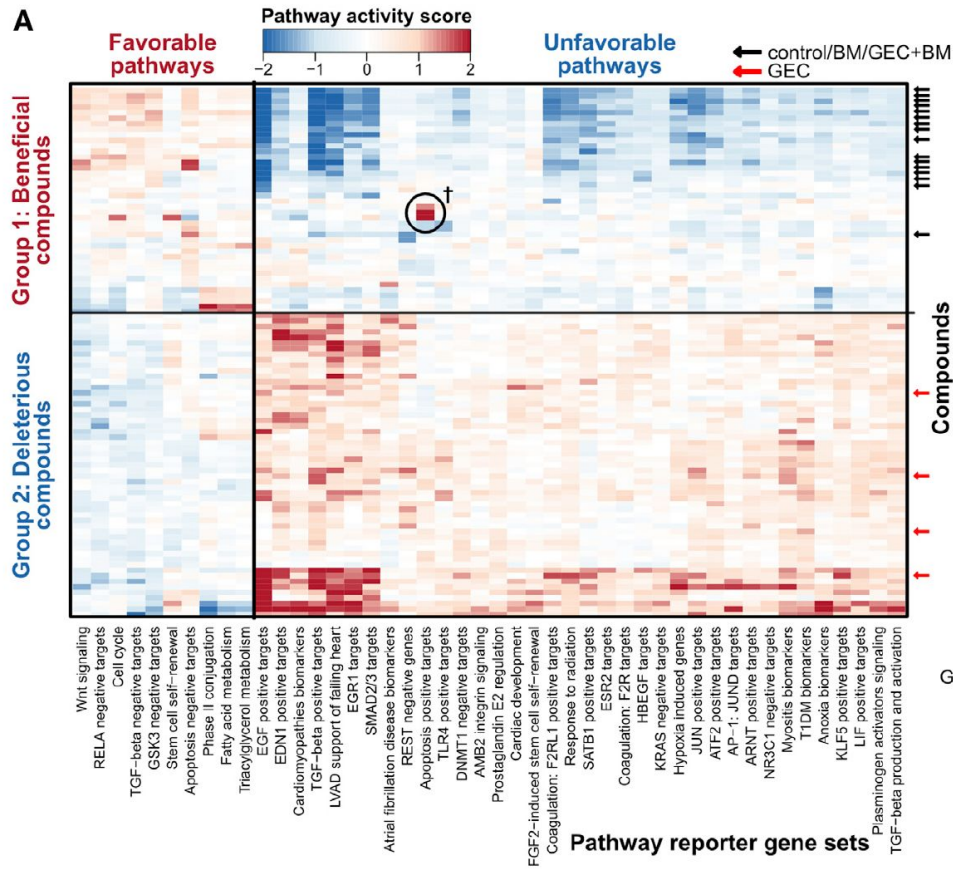
Input: (1) a differential gene expression profile; (2) a set of gene-sets $\{G\}$, each a set of genes.

Output: a ranked list of the input gene-sets by *enrichment*.

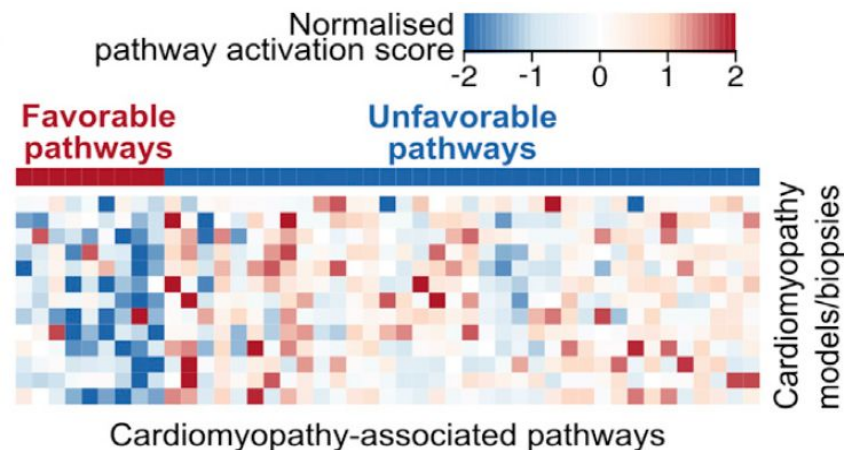
Gene expression as screening readout



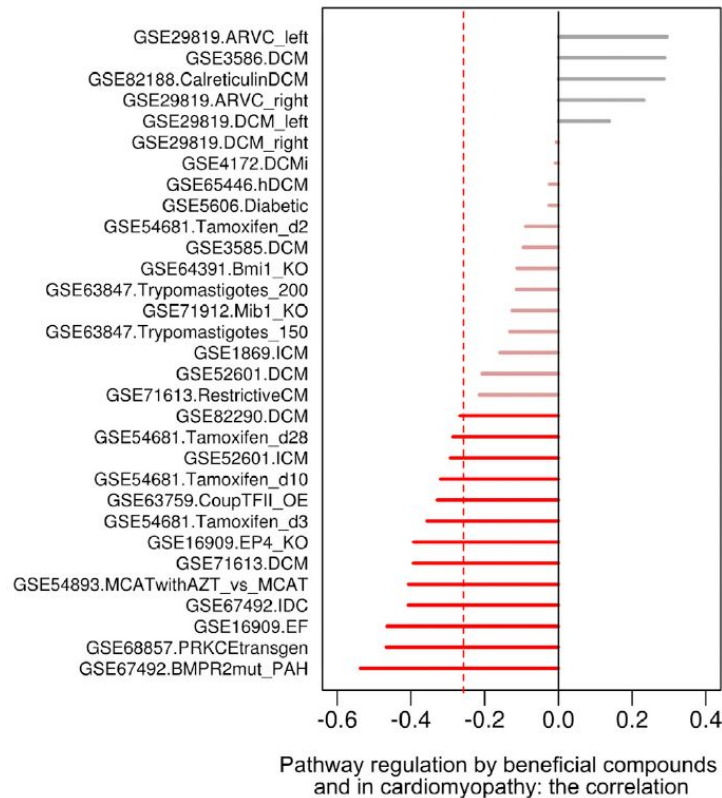
Differential gene expression profiles are molecular snapshots of drugs' action in the cell.



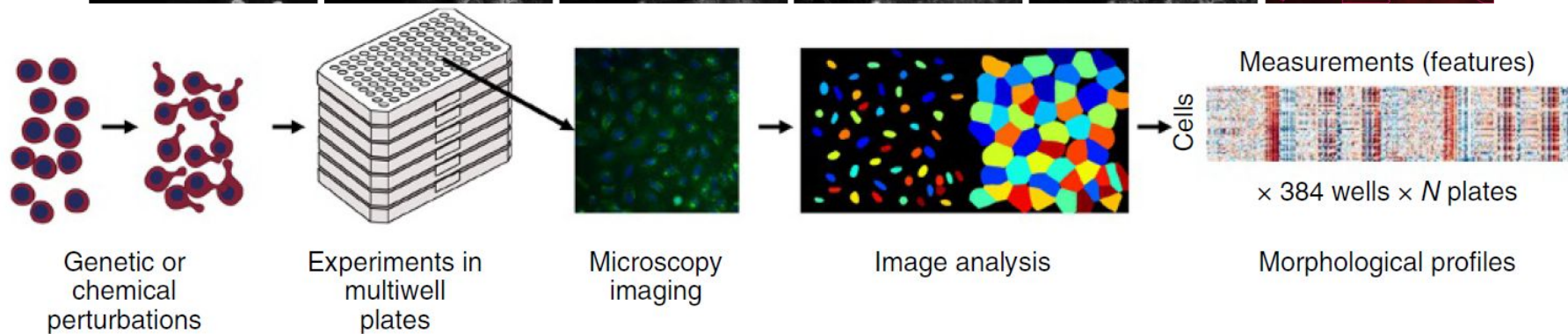
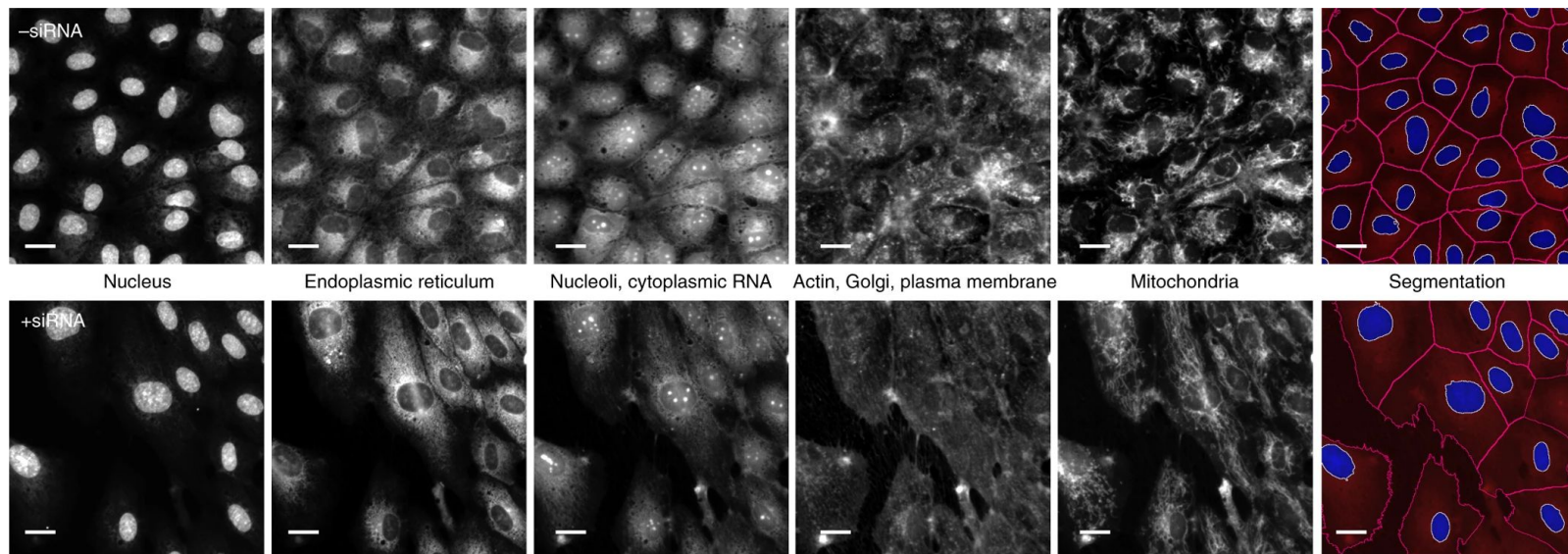
Gene expression from patient and animal models help compound selection



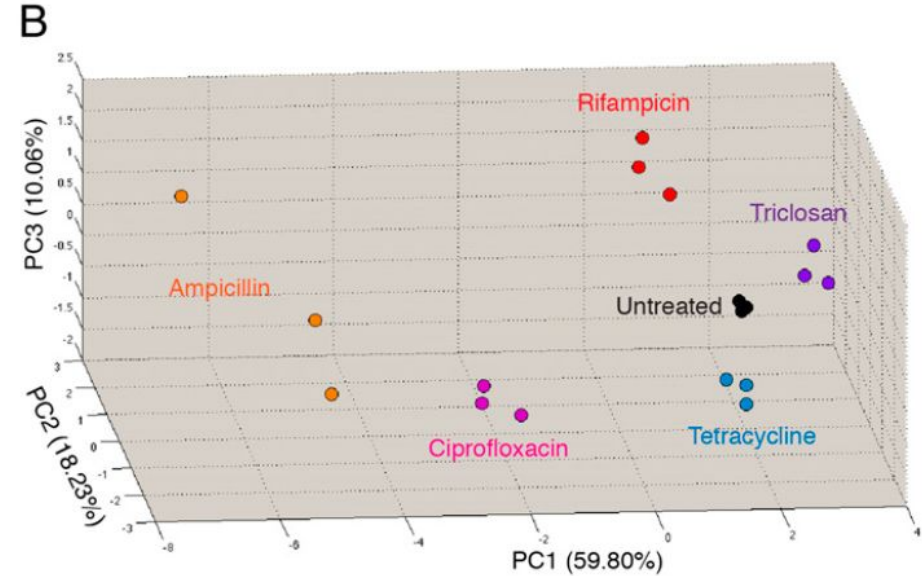
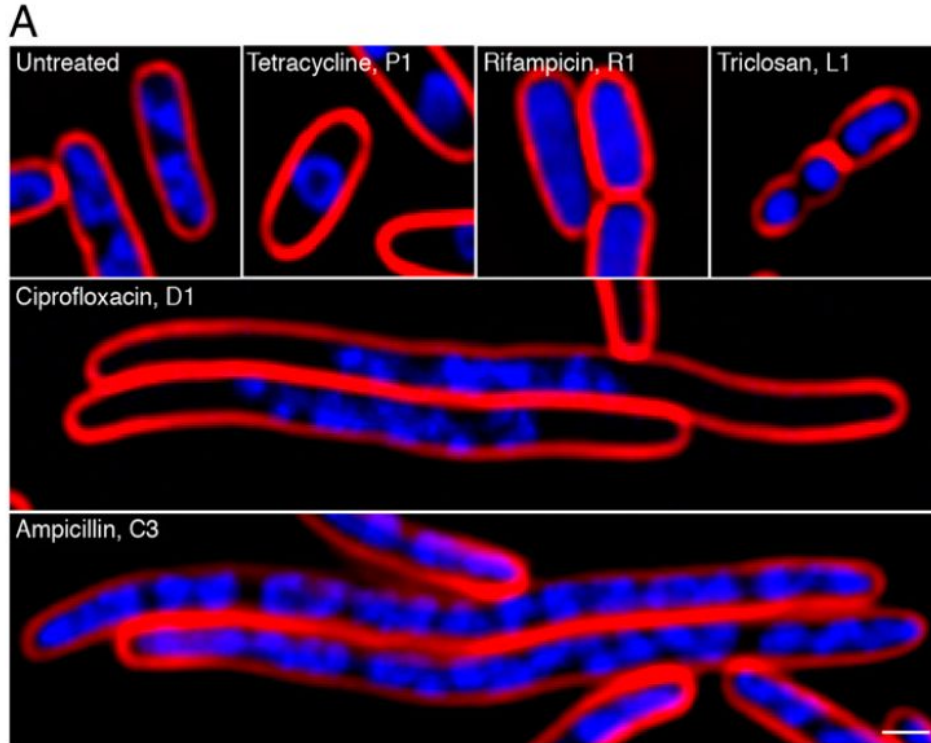
We can prioritise molecules that reverse disease-induced changes.



Morphology as screening readout



Cytological profiling for antibiotics discovery



P: Protein translation inhibitors

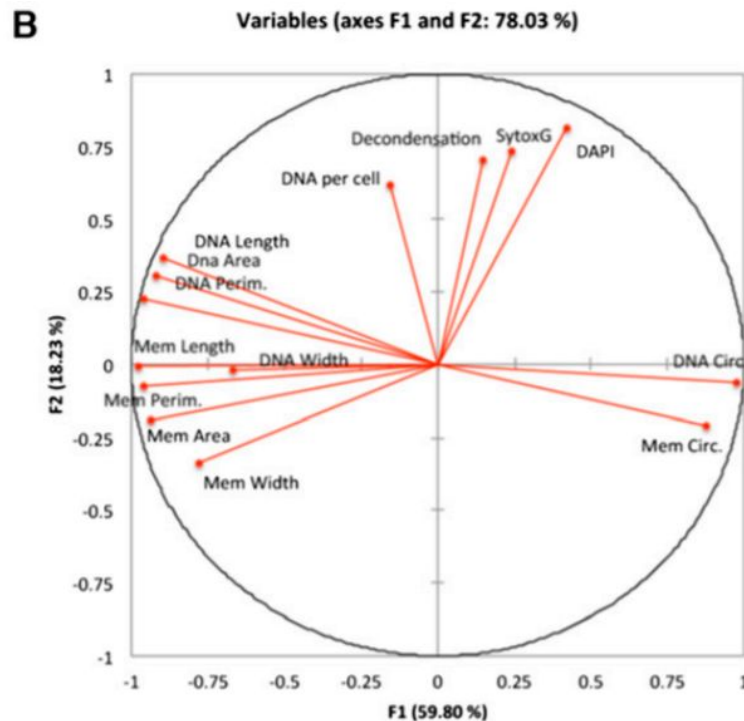
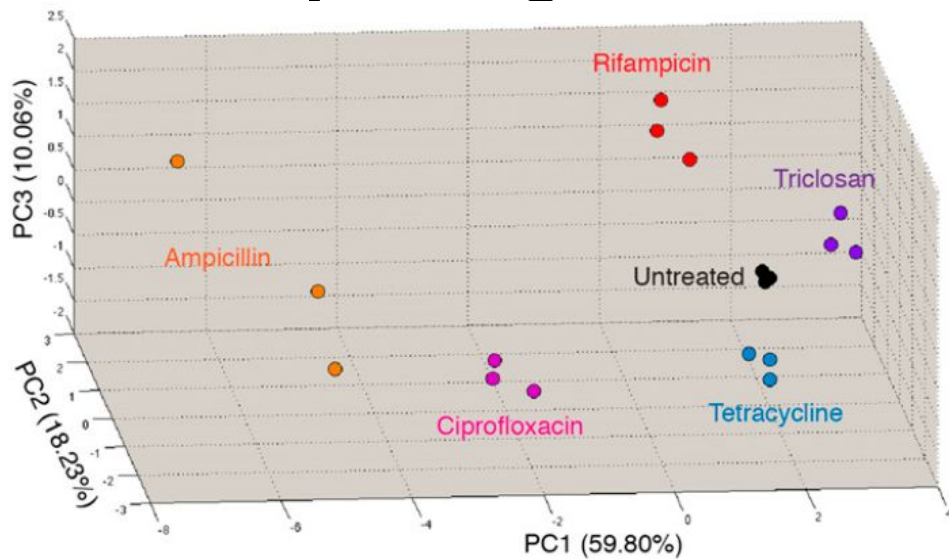
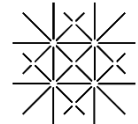
R: RNA transcription inhibitors

D: DNA replication inhibitors

L: Lipid biosynthesis inhibitors

C: Cell-wall synthesis inhibitors (peptidoglycan)

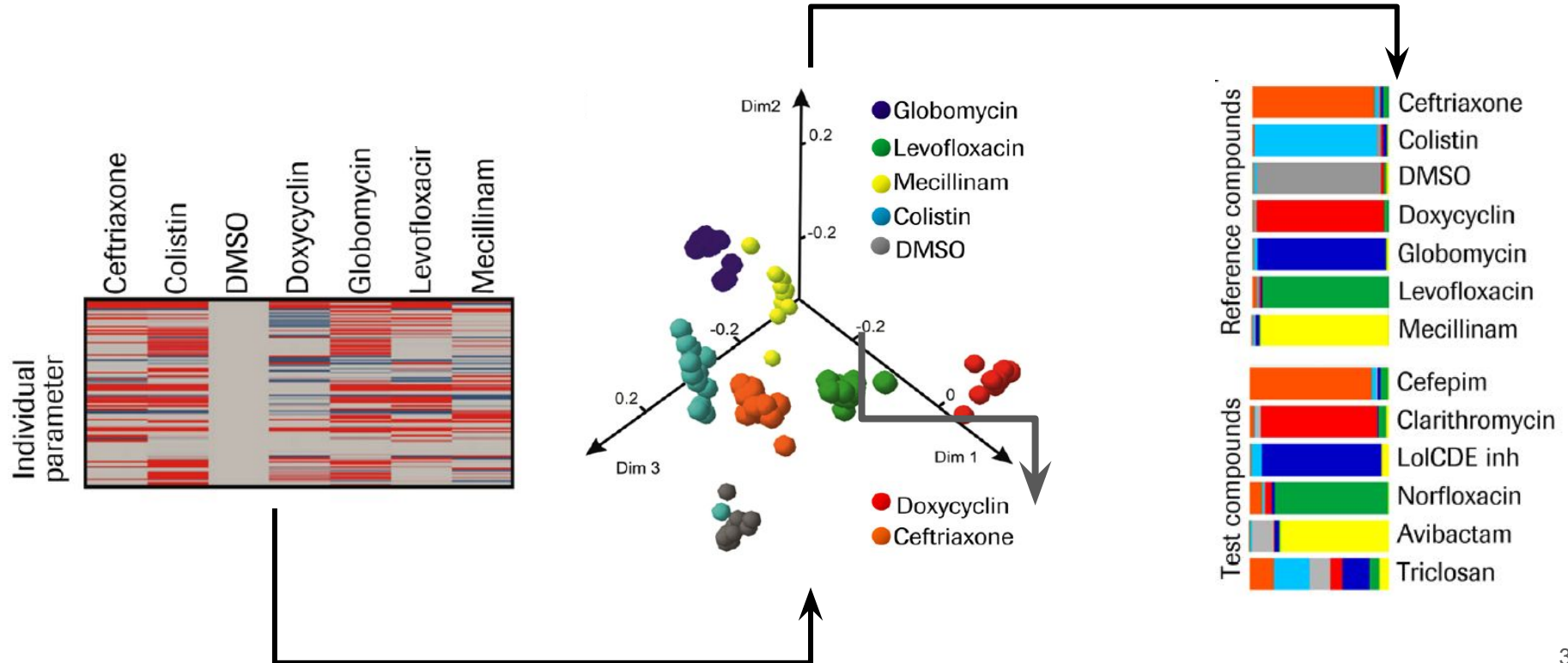
Principal components are linear combination of morphological features



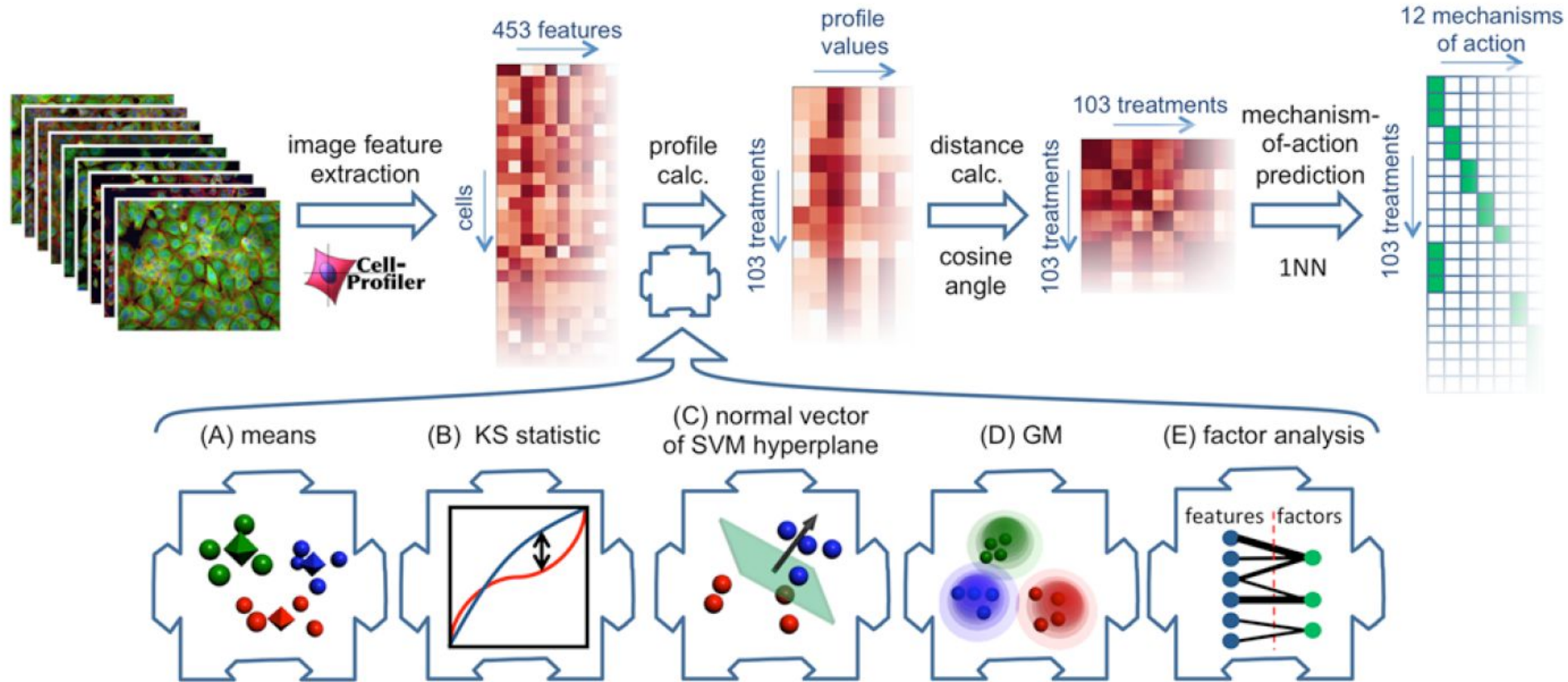
Membrane area, μm^2 DNA area, μm^2 Membrane perimeter, μm DNA perimeter, μm Membrane length, μm DNA length, μm No. of nucleoids per cell

Membrane width, μm DNA width, μm Membrane circularity DNA circularity SytoxG intensity DAPI intensity Decondensation

Morphology classifies compounds by MoA



Comparison of computational methods



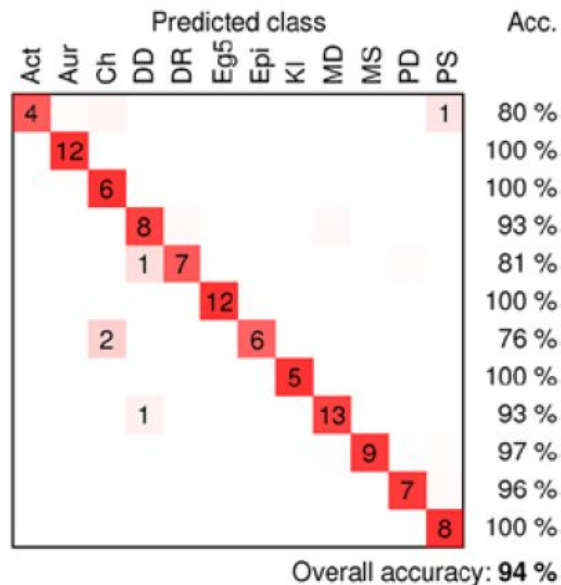
Do the benchmark and use Occam's Razor

Table 1. Accuracies for classifying compound treatments into mechanisms of action.

Method	Accuracy, %
Means	83
KS statistic	83
Normal vector to support-vector machine hyperplane	81
With recursive feature elimination	64
Distribution over Gaussian mixture components	83
Factor analysis + means	94

True mechanistic class

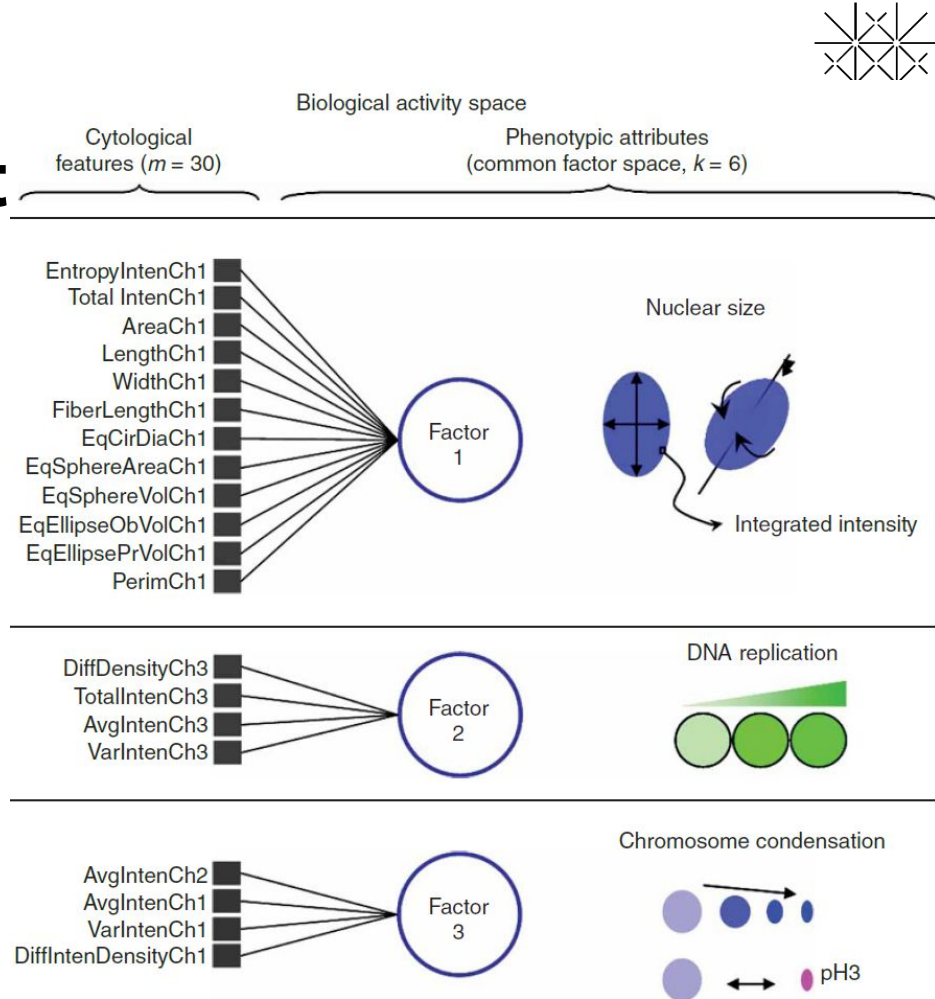
Actin disruptors	Act
Aurora kinase inhibitors	Aur
Cholesterol-lowering	Ch
DNA damage	DD
DNA replication	DR
Eg5 inhibitors	Eg5
Epithelial	Epi
Kinase inhibitors	KI
Microtubule destabilizers	MD
Microtubule stabilizers	MS
Protein degradation	PD
Protein synthesis	PS



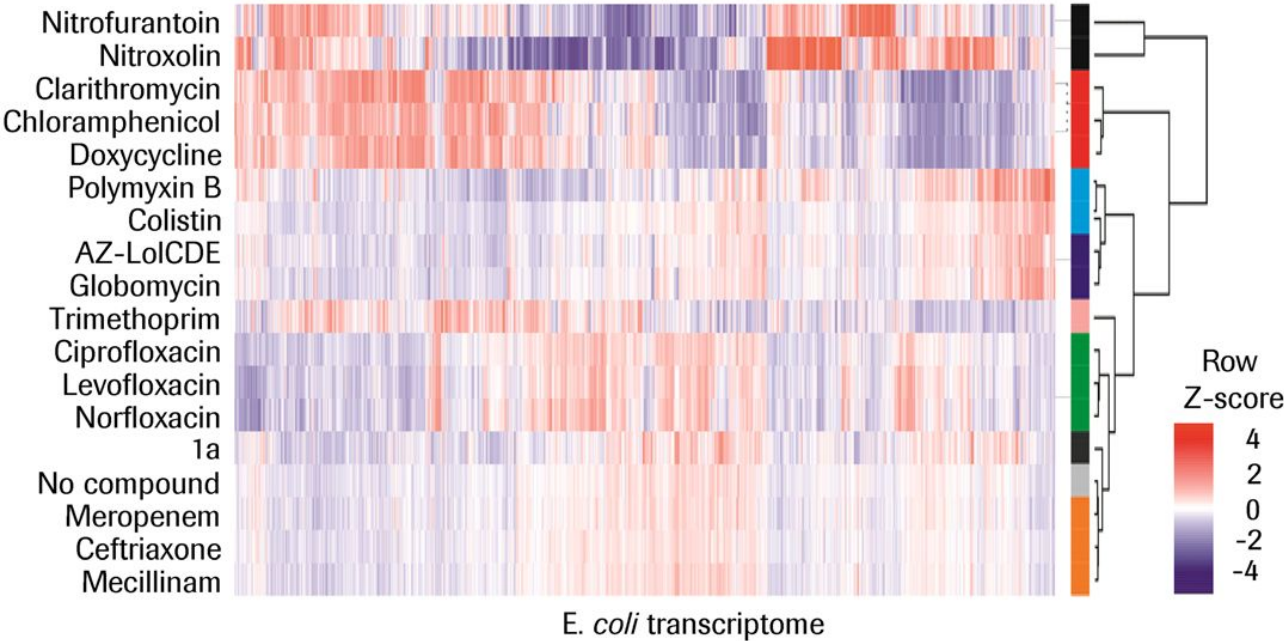
A possible explanation for the success of latent variable models

$$\begin{matrix} \text{Cells} \\ \begin{pmatrix} x_{11} & \dots & x_{1m} \\ \vdots & \ddots & \vdots \\ x_{n1} & \dots & x_{nm} \end{pmatrix} \\ \text{Cytological features} \end{matrix} = X_{nm} = \underbrace{\sum_{i=1}^k L_{ni} F_{im}}_{k\text{-factor space}} + \varepsilon_{nm}$$

A common latent factor model



Morphology and gene expression used jointly

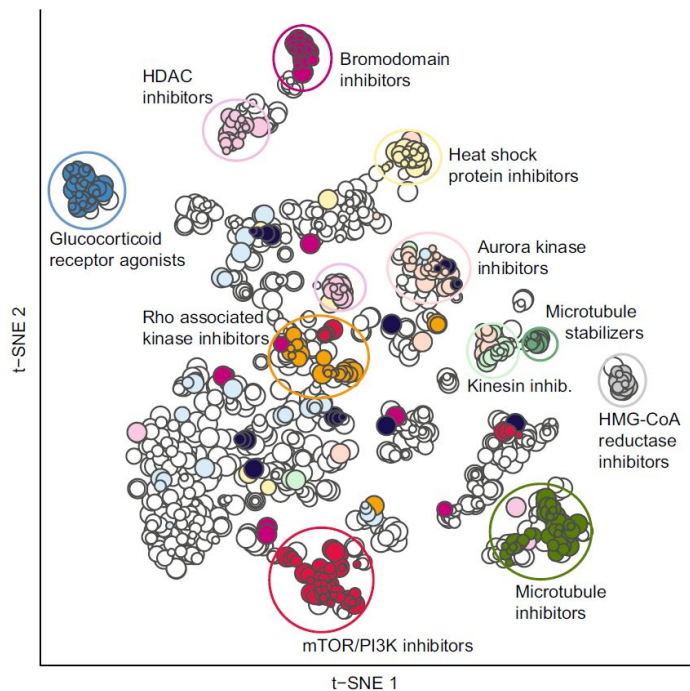


Gene-set
enrichment
analysis

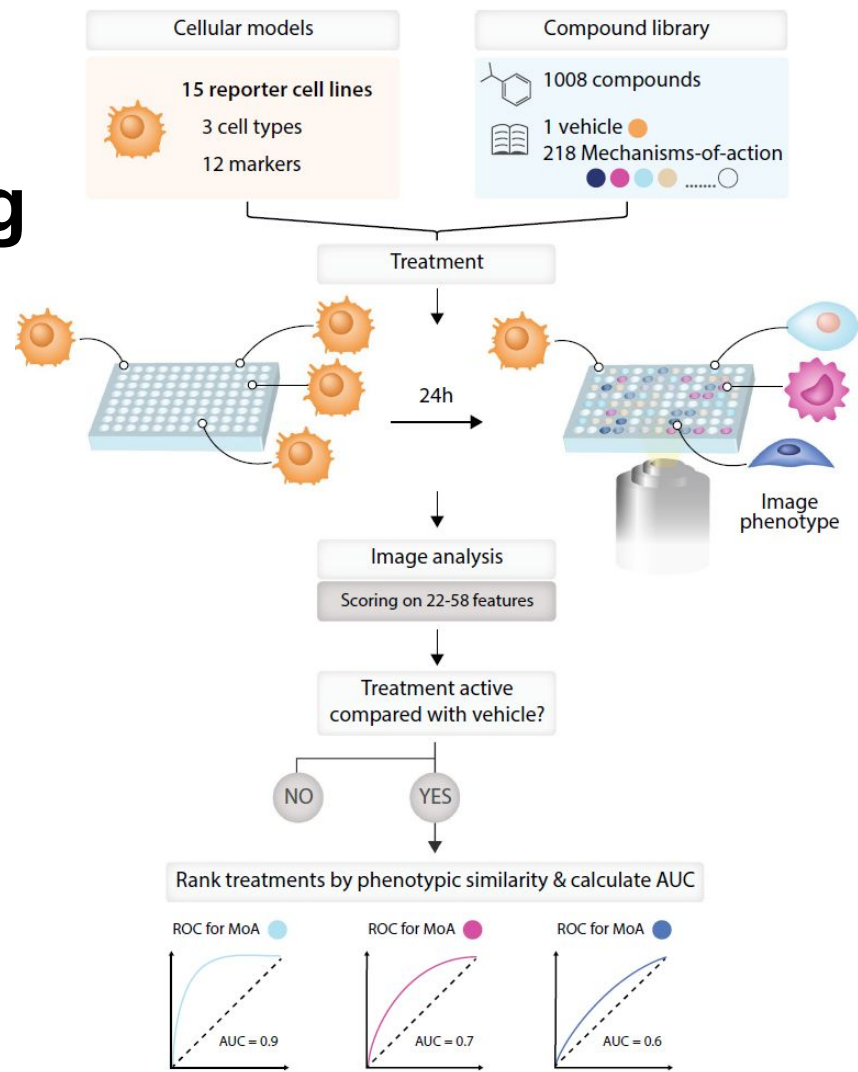
Reporter
assays

Pathway-
Phenotype
associations

A multi-cell-type, 1008-compound screening by Cox *et al.* (2020)



- 0.3 μ M
- 1 μ M
- 3 μ M
- 9 μ M
- ABL1 inhibitor
- Aurora kinase inhibitor
- Bromodomain inhibitor
- Glucocorticoid receptor agonist
- HDAC inhibitor
- Heat shock protein inhibitor
- HMG-CoA reductase inhibitor
- Kinesin inhibitor
- Microtubule inhibitor
- Microtubule stabilizer
- mTOR and/or PI3K inhibitor
- Rho associated kinase inhibitor
- VEGFR family inhibitor
- (Other)



Conclusions

- Gene expression and image-based profiling can be used individually or jointly for phenotypic screening;
- Integration of biological knowledge, high-throughput data, and statistical modelling empowers phenotypic drug discovery.

References

1. Swinney, David C., and Jason Anthony. 2011. "How Were New Medicines Discovered?" *Nature Reviews Drug Discovery* 10 (7): 507–19. <https://doi.org/10.1038/nrd3480>.
2. Marx, Uwe, Tommy B. Andersson, Anthony Bahinski, Mario Beilmann, Sonja Beken, Flemming R. Cassee, Murat Cirit, et al. 2016. "Biology-Inspired Microphysiological System Approaches to Solve the Prediction Dilemma of Substance Testing." *ALTEX - Alternatives to Animal Experimentation* 33 (3): 272–321. <https://doi.org/10.14573/altex.1603161>.
3. Dickey, Seth W., Gordon Y. C. Cheung, and Michael Otto. 2017. "Different Drugs for Bad Bugs: Antivirulence Strategies in the Age of Antibiotic Resistance." *Nature Reviews Drug Discovery* 16 (7): 457–71. <https://doi.org/10.1038/nrd.2017.23>.
4. Lewis, Kim. 2013. "Platforms for Antibiotic Discovery." *Nature Reviews Drug Discovery* 12 (5): 371–87. <https://doi.org/10.1038/nrd3975>.
5. Warner, Katherine Deigan, Christine E. Hajdin, and Kevin M. Weeks. 2018. "Principles for Targeting RNA with Drug-like Small Molecules." *Nature Reviews Drug Discovery* 17 (8): 547–58. <https://doi.org/10.1038/nrd.2018.93>.
6. Berg, Ellen L. 2021. "The Future of Phenotypic Drug Discovery." *Cell Chemical Biology* 28 (3): 424–30. <https://doi.org/10.1016/j.chembiol.2021.01.010>.
7. Chin, Marcus Y., Jether Amos Espinosa, Grace Pohan, Sarine Markossian, and Michelle R. Arkin. 2021. "Reimagining Dots and Dashes: Visualizing Structure and Function of Organelles for High-Content Imaging Analysis." *Cell Chemical Biology* 28 (3): 320–37. <https://doi.org/10.1016/j.chembiol.2021.01.016>.
8. Conway, Louis P., Weichao Li, and Christopher G. Parker. 2021. "Chemoproteomic-Enabled Phenotypic Screening." *Cell Chemical Biology* 28 (3): 371–93. <https://doi.org/10.1016/j.chembiol.2021.01.012>.
9. Dahlin, Jayme L., Douglas S. Auld, Ina Rothenaigner, Steve Haney, Jonathan Z. Sexton, J. Willem M. Nissink, Jarrod Walsh, et al. 2021. "Nuisance Compounds in Cellular Assays." *Cell Chemical Biology* 28 (3): 356–70. <https://doi.org/10.1016/j.chembiol.2021.01.021>.
10. Ha, Jaeyoung, Hankum Park, Jongmin Park, and Seung Bum Park. 2021. "Recent Advances in Identifying Protein Targets in Drug Discovery." *Cell Chemical Biology* 28 (3): 394–423. <https://doi.org/10.1016/j.chembiol.2020.12.001>.

References (continued)

11. Hsu, Ku-Lung. 2021. "Shining a Light on Phenotypic Drug Discovery." *Cell Chemical Biology* 28 (2): 115–17. <https://doi.org/10.1016/j.chembiol.2021.01.020>.
12. Hughes, Rebecca E., Richard J. R. Elliott, John C. Dawson, and Neil O. Carragher. 2021. "High-Content Phenotypic and Pathway Profiling to Advance Drug Discovery in Diseases of Unmet Need." *Cell Chemical Biology* 28 (3): 338–55. <https://doi.org/10.1016/j.chembiol.2021.02.015>.
13. Vandana, J. Jeya, Lauretta A. Lacko, and Shuibing Chen. 2021. "Phenotypic Technologies in Stem Cell Biology." *Cell Chemical Biology* 28 (3): 257–70. <https://doi.org/10.1016/j.chembiol.2021.02.001>.
14. Ziegler, Slava, Sonja Sievers, and Herbert Waldmann. 2021. "Morphological Profiling of Small Molecules." *Cell Chemical Biology* 28 (3): 300–319. <https://doi.org/10.1016/j.chembiol.2021.02.012>.
15. Moffat, John G., Fabien Vincent, Jonathan A. Lee, Jörg Eder, and Marco Prunotto. 2017. "Opportunities and Challenges in Phenotypic Drug Discovery: An Industry Perspective." *Nature Reviews Drug Discovery* 16 (8): 531–43. <https://doi.org/10.1038/nrd.2017.111>.
16. Drawnel, Faye Marie, Jitao David Zhang, Erich Küng, Natsuyo Aoyama, Fethallah Benmansour, Andrea Araujo Del Rosario, Sannah Jensen Zoffmann, et al. 2017. "Molecular Phenotyping Combines Molecular Information, Biological Relevance, and Patient Data to Improve Productivity of Early Drug Discovery." *Cell Chemical Biology* 18 (24(5)): 624–34. <https://doi.org/10.1016/j.chembiol.2017.03.016>.
17. Roudnicky, Filip, Jitao David Zhang, Bo Kyoung Kim, Nikhil J. Pandya, Yanjun Lan, Lisa Sach-Peltason, Heloise Ragelle, et al. 2020. "Inducers of the Endothelial Cell Barrier Identified through Chemogenomic Screening in Genome-Edited HPSC-Endothelial Cells." *Proceedings of the National Academy of Sciences*, August. <https://doi.org/10.1073/pnas.1911532117>.
18. Zoffmann, Sannah, Maarten Vercruysse, Fethallah Benmansour, Andreas Maunz, Luise Wolf, Rita Blum Marti, Tobias Heckel, et al. 2019. "Machine Learning-Powered Antibiotics Phenotypic Drug Discovery." *Scientific Reports* 9 (1): 1–14. <https://doi.org/10.1038/s41598-019-39387-9>.
19. Nichols, Robert J., Saunak Sen, Yoe Jin Choo, Pedro Beltrao, Matylda Zietek, Rachna Chaba, Sueyoung Lee, et al. 2011. "Phenotypic Landscape of a Bacterial Cell." *Cell* 144 (1): 143–56. <https://doi.org/10.1016/j.cell.2010.11.052>.
20. Smith, Kevin, Filippo Piccinini, Tamas Balassa, Krisztian Koos, Tivadar Danko, Hossein Azizpour, and Peter Horvath. 2018. "Phenotypic Image Analysis Software Tools for Exploring and Understanding Big Image Data from Cell-Based Assays." *Cell Systems* 6 (6): 636–53. <https://doi.org/10.1016/j.cels.2018.06.001>.

References (continued)

21. Scheeder, Christian, Florian Heigwer, and Michael Boutros. 2018. "Machine Learning and Image-Based Profiling in Drug Discovery." *Current Opinion in Systems Biology, Pharmacology and drug discovery*, 10 (August): 43–52. <https://doi.org/10.1016/j.coisb.2018.05.004>.
22. Schirle, Markus, and Jeremy L. Jenkins. 2016. "Identifying Compound Efficacy Targets in Phenotypic Drug Discovery." *Drug Discovery Today* 21 (1): 82–89. <https://doi.org/10.1016/j.drudis.2015.08.001>.
23. Wilkinson, Isabel V. L., Georg C. Terstappen, and Angela J. Russell. 2020. "Combining Experimental Strategies for Successful Target Deconvolution." *Drug Discovery Today* 25 (11): 1998–2005. <https://doi.org/10.1016/j.drudis.2020.09.016>.
24. Comess, Kenneth M., Shaun M. McLoughlin, Jon A. Oyer, Paul L. Richardson, Henning Stöckmann, Anil Vasudevan, and Scott E. Warder. 2018. "Emerging Approaches for the Identification of Protein Targets of Small Molecules - A Practitioners' Perspective." *Journal of Medicinal Chemistry* 61 (19): 8504–35. <https://doi.org/10.1021/acs.jmedchem.7b01921>.
25. Sidders, Ben, Anna Karlsson, Linda Kitching, Rubben Torella, Paul Karila, and Anne Phelan. 2018. "Network-Based Drug Discovery: Coupling Network Pharmacology with Phenotypic Screening for Neuronal Excitability." *Journal of Molecular Biology, Theory and Application of Network Biology Toward Precision Medicine*, 430 (18, Part A): 3005–15. <https://doi.org/10.1016/j.jmb.2018.07.016>.
26. Aulner, Nathalie, Anne Danckaert, JongEun Ihm, David Shum, and Spencer L. Shorte. 2019. "Next-Generation Phenotypic Screening in Early Drug Discovery for Infectious Diseases." *Trends in Parasitology* 35 (7): 559–70. <https://doi.org/10.1016/j.pt.2019.05.004>.
27. Boess, Franziska, Barbara Lenz, Juergen Funk, Urs Niederhauser, Simon Bassett, Jitao David Zhang, Thomas Singer, and Adrian B. Roth. 2017. "Use of Early Phenotypic in Vivo Markers to Assess Human Relevance of an Unusual Rodent Non-Genotoxic Carcinogen in Vitro." *Toxicology* 379 (March): 48–61. <https://doi.org/10.1016/j.tox.2017.01.018>.
28. Feng, Yan, Timothy J. Mitchison, Andreas Bender, Daniel W. Young, and John A. Tallarico. 2009. "Multi-Parameter Phenotypic Profiling: Using Cellular Effects to Characterize Small-Molecule Compounds." *Nature Reviews Drug Discovery* 8 (7): 567–78. <https://doi.org/10.1038/nrd2876>.
29. Jones, Lyn H., and Mark E. Bunnage. 2017. "Applications of Chemogenomic Library Screening in Drug Discovery." *Nature Reviews Drug Discovery* 16 (January): 285–296. <https://doi.org/10.1038/nrd.2016.244>.
30. Kulesa, Anthony, Jared Kehe, Juan E. Hurtado, Prianca Tawde, and Paul C. Blainey. 2018. "Combinatorial Drug Discovery in Nanoliter Droplets." *Proceedings of the National Academy of Sciences* 115 (26): 6685–90. <https://doi.org/10.1073/pnas.1802233115>.

References (continued)

31. Vlachogiannis, Georgios, Somaieh Hedayat, Alexandra Vatsiou, Yann Jamin, Javier Fernández-Mateos, Khurum Khan, Andrea Lampis, et al. 2018. "Patient-Derived Organoids Model Treatment Response of Metastatic Gastrointestinal Cancers." *Science* 359 (6378): 920–26.
<https://doi.org/10.1126/science.aao2774>.
32. Wawer, Mathias J., Kejie Li, Sigrun M. Gustafsdottir, Vebjorn Ljosa, Nicole E. Bodycombe, Melissa A. Marton, Katherine L. Sokolnicki, et al. 2014. "Toward Performance-Diverse Small-Molecule Libraries for Cell-Based Phenotypic Screening Using Multiplexed High-Dimensional Profiling." *Proceedings of the National Academy of Sciences* 111 (30): 10911–16. <https://doi.org/10.1073/pnas.1410933111>.
33. Zhang, Jitao David, Erich Küng, Franziska Boess, Ulrich Certa, and Martin Ebeling. 2015. "Pathway Reporter Genes Define Molecular Phenotypes of Human Cells." *BMC Genomics* 16: 342. <https://doi.org/10.1186/s12864-015-1532-2>.
34. Antolin, Albert A., Joseph E. Tym, Angeliki Komianou, Ian Collins, Paul Workman, and Bissan Al-Lazikani. 200017. "Objective, Quantitative, Data-Driven Assessment of Chemical Probes." *Cell Chemical Biology* 0 (0). <https://doi.org/10.1016/j.chembiol.2017.11.4>.
35. Finotello, Francesca, and Barbara Di Camillo. 2015. "Measuring Differential Gene Expression with RNA-Seq: Challenges and Strategies for Data Analysis." *Briefings in Functional Genomics* 14 (2): 130–42. <https://doi.org/10.1093/bfpg/elu035>.
36. Anders, Simon, and Wolfgang Huber. 2010. "Differential Expression Analysis for Sequence Count Data." *Genome Biology* 11 (10): R106.
<https://doi.org/10.1186/gb-2010-11-10-r106>.
37. Robinson, Mark D., and Gordon K. Smyth. 2007. "Moderated Statistical Tests for Assessing Differences in Tag Abundance." *Bioinformatics* 23 (21): 2881–87.
<https://doi.org/10.1093/bioinformatics/btm453>.
38. Jesse Lipp, *Why sequencing data is modeled as negative binomial*,
<https://bioramble.wordpress.com/2016/01/30/why-sequencing-data-is-modeled-as-negative-binomial/>
39. Stanton, Benjamin Z., Emma J. Chory, and Gerald R. Crabtree. 2018. "Chemically Induced Proximity in Biology and Medicine." *Science* 359 (6380): [eaa05902](https://doi.org/10.1126/science.aao5902).
<https://doi.org/10.1126/science.aao5902>.
40. Tom Gregory, <https://www.quora.com/Is-Gini-coefficient-outdated>

References (continued)

41. [EBI online tutorial of Reactome](#), DOI: 10.6019/TOL.Reactome_exbp-t.2017.00001.1
42. Frey, Brendan J., and Delbert Dueck. 2007. "Clustering by Passing Messages Between Data Points." *Science* 315 (5814): 972–76.
<https://doi.org/10.1126/science.1136800>.
43. RNA-seq: <https://en.wikipedia.org/wiki/RNA-Seq>, in particular the [figure](#) by Thomas Shafee (CC BY 4.0)
44. [Affinity Propagation algorithm visualized](#)
45. Bray, Mark-Anthony, Shantanu Singh, Han Han, Chadwick T. Davis, Blake Borgeson, Cathy Hartland, Maria Kost-Alimova, Sigrun M. Gustafsdottir, Christopher C. Gibson, and Anne E. Carpenter. 2016. "Cell Painting, a High-Content Image-Based Assay for Morphological Profiling Using Multiplexed Fluorescent Dyes." *Nature Protocols* 11 (9): 1757–74. <https://doi.org/10.1038/nprot.2016.105>.
46. Nonejuie, Poochit, Michael Burkart, Kit Pogliano, and Joe Pogliano. 2013. "Bacterial Cytological Profiling Rapidly Identifies the Cellular Pathways Targeted by Antibacterial Molecules." *Proceedings of the National Academy of Sciences* 110 (40): 16169–74. <https://doi.org/10.1073/pnas.1311066110>.
47. Ljosa, Vebjorn, Peter D. Caie, Rob ter Horst, Katherine L. Sokolnicki, Emma L. Jenkins, Sandeep Daya, Mark E. Roberts, et al. 2013. "Comparison of Methods for Image-Based Profiling of Cellular Morphological Responses to Small-Molecule Treatment." *Journal of Biomolecular Screening* 18 (10): 1321–29.
<https://doi.org/10.1177/1087057113503553>.
48. Young, Daniel W., Andreas Bender, Jonathan Hoyt, Elizabeth McWhinnie, Gung-Wei Chirn, Charles Y. Tao, John A. Tallarico, et al. 2008. "Integrating High-Content Screening and Ligand-Target Prediction to Identify Mechanism of Action." *Nature Chemical Biology* 4 (1): 59–68.
<https://doi.org/10.1038/nchembio.2007.53>.
49. Cox, Michael J., Steffen Jaensch, Jelle Van de Waeter, Laure Cougnaud, Daan Seynaeve, Soulaïman Benalla, Seong Joo Koo, et al. 2020. "Tales of 1,008 Small Molecules: Phenomic Profiling through Live-Cell Imaging in a Panel of Reporter Cell Lines." *Scientific Reports* 10 (1): 13262.
<https://doi.org/10.1038/s41598-020-69354-8>.
50. Stark, Rory, Marta Grzelak, and James Hadfield. 2019. "RNA Sequencing: The Teenage Years." *Nature Reviews Genetics*, July, 1–26.
<https://doi.org/10.1038/s41576-019-0150-2>.